Searching for candidate speciation genes using a proteomic approach: seminal proteins in field crickets

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In many animals, male seminal proteins influence gamete interactions and fertilization ability and are probably involved in barriers to gene flow between diverging lineages. Here we use a proteomic approach to identify seminal proteins that are transferred to females during copulation and that may be involved in fertilization barriers between two hybridizing field crickets (Gryllus firmus and Gryllus pennsylvanicus). Analyses of patterns of divergence suggest that much of the field cricket genome has remained undifferentiated following the evolution of reproductive isolation. By contrast, seminal protein genes are highly differentiated. Tests of selection reveal that positive selection is likely to be responsible for patterns of differentiation. Together, our observations suggest that some of the loci encoding seminal proteins may indeed play a role in fertilization barriers in field crickets.

Keywords: reproductive isolation; fertilization barriers; gene genealogies; selection; Gryllus

1. INTRODUCTION

Identifying genetic changes that contribute to reproductive isolation is one of the central problems in evolutionary biology. Although almost all of the genes of an organism potentially can contribute to intraspecific variation and population differentiation, only relatively few genes, so-called ‘speciation’ or ‘barrier’ genes (Wu 2001; Noor & Feder 2006), are presumably involved in barriers to gene flow between diverging lineages. Historically, studies of speciation have focused primarily on the reduced fitness of hybrids (postzygotic isolation) and on premating barriers to gene exchange, especially behavioural isolation (Coyne & Orr 2004). More recently, attention has been drawn to fertilization barriers as components of reproductive isolation and speciation. For example, in several groups of marine external fertilizers (sea urchins, abalones, turban snails and mussels) gamete recognition proteins have been identified and shown to be evolving rapidly under directional selection (Metz & Palumbi 1996; Hellberg & Vacquier 1999; Yang et al. 2000; Swanson & Vacquier 2002; Riginos et al. 2006).

In many animals fertilization is internal, and zygote formation is mediated/facilitated not only by sperm and egg proteins but also by proteins present in the seminal fluid (seminal proteins). In insects and primates, a subset of seminal proteins is among the most rapidly evolving proteins and, like gamete recognition proteins in marine invertebrates, exhibit a clear signature of positive selection (Clark et al. 2006). The rapid adaptive evolution of these proteins (Clark & Swanson 2005; Mueller et al. 2005; Andrés et al. 2006) may be an important component of reproductive isolation during the early stages of the speciation process (Andrés & Arntqvist 2001; Coyne & Orr 2004).

Locii encoding seminal proteins are likely to be involved in the barriers that generate new species because these genes determine traits that influence sperm activation, gamete interactions and ovulation (Peitz 1988; Tram & Wollner 1999; Viscuso et al. 2001; Fry & Wilkinson 2004). In polyandrous species, seminal proteins play a key role in postmating sexual selection (Fiúmera et al. 2005). Postcopulatory competition for the fertilization of female eggs (sperm competition and selective fertilization) may lead to rapid coevolution between seminal proteins (those transferred from males to females during copulation) and proteins of the female reproductive tract. Independent episodes of rapid coevolution (e.g. in allopatric populations) could lead to reproductive divergence and ultimately to speciation (Howard 1999; Andrés & Arntqvist 2001; Coyne & Orr 2004; Fricke et al. 2006).

Genealogical analyses and tests of selection can be used to assess whether genes encoding seminal proteins are potentially involved in reproductive isolation. Because genomes are genealogical or historical mosaics (Wu 2001), gene genealogies for closely related species vary with genomic region (Ting et al. 2000; Machado & Hey 2003; Dopman et al. 2005). Regions of the genome contributing to adaptive divergence and reproductive isolation will diverge more rapidly owing to the selective sorting of ancestral polymorphisms and/or the selective disadvantage of introgressed alleles (Putman et al. 2007). Hence, gene genealogies for loci that have been subject to recent selective sweeps, including those that contribute to reproductive isolation, are more likely to reveal closely related species as differentiated or exclusive (i.e. monophyletic) groups; at most other loci, shared ancestral polymorphism will persist and/or introgression will erase differences that are accumulated in allopatry (Ting et al. 2000).

The closely related field crickets Gryllus firmus and Gryllus pennsylvanicus, which come into contact in a well-characterized hybrid zone in eastern North America.
(Harrison & Bogdanowicz 1997; Ross & Harrison 2002), provide an important model system for investigating the origin of barriers to gene exchange. The net mtDNA sequence divergence between this species pair is less than 0.5% (Willett et al. 1997), and surveys of variation in allozymes and nuclear gene introns have failed to reveal genome regions for which the two species exhibit fixed differences or are exclusive groups (Harrison 1979; Broughton & Harrison 2003). The estimated time since divergence for G. firmus and G. pennsylvanicus is only 0.1Nₑ; therefore, we expect ancestral polymorphisms to segregate in most regions of the genome (Broughton & Harrison 2003). Loci for which the two species are highly differentiated or exclusive groups probably mark genome regions that have experienced recent selective sweeps and/or harbour barrier genes.

Field and laboratory studies of the two cricket species (Harrison 1983, 1985, 1986; Harrison & Rand 1989) have identified both pre- and postmating barriers to gene exchange. Premating barriers include ecological (habitat) isolation, temporal isolation and mate choice. However, premating barriers are not complete, and adults of the two species, together with individuals of mixed ancestry, are found together at hybrid zone localities. The only known postmating barrier is a one-way incompatibility between G. firmus females and G. pennsylvanicus males. That is, heterospecific males do not trigger normal oviposition behaviour in G. firmus females, and the eggs produced, which do not develop, are indistinguishable from unfertilized eggs (Harrison 1983; L. S. Maroja, J. A. Andrés & R. G. Harrison 2008, unpublished data). Furthermore, an evolutionarily expressed sequence tag screen of the male cricket accessory gland has identified a set of genes encoding secreted proteins that show male-biased gene expression (Andrés et al. 2006; Braswell et al. 2006). These secreted proteins, some of which exhibit rapid evolution and evidence of positive selection, are putative seminal proteins.

Here we use a proteomic approach to provide unambiguous identification of genes encoding proteins that are transferred from male to female crickets during copulation. The approach relies on comparison of peptide sequences from tryptic digests of spermatophore homogenates with peptide sequences generated in silico from a translation of cricket accessory gland expressed sequence tag (EST) database. Matching of peptides within the spermatophore with those generated in silico allows definitive identification of genes that encode seminal proteins. Direct identification of seminal proteins is more reliable than a purely bioinformatics approach and avoids the possibility of missing rapidly evolving genes. The goal is to target a particular class of genes that may well be subject to positive selection and also may play a role in reproductive isolation. We then compare genealogies for genes encoding seminal proteins with genealogies for genes that are not related to reproduction. Our observations suggest that loci encoding seminal proteins may indeed play a role in barriers to gene exchange in field crickets.

2. MATERIAL AND METHODS

(a) Protein sample preparation

Two independent samples of male spermatophores (n = 6 males per sample), the proteinaceous capsule containing the ejaculate, were harvested directly from G. pennsylvanicus males and homogenized in 100 μl of ice-cold phosphate buffer. The two independent samples were centrifuged (14 000g at 4°C) to separate seminal fluid from most of the sperm and spermatophore debris. Samples were stored at −80°C, and aliquots of each were sent to the Genome BC Proteomics Centre (University of Victoria, Canada), where proteomic analyses were carried out as described below.

Seminal fluid proteins were solubilized by adding 9.5 M urea, 50 mM NH₄HCO₃ and 0.2% SDS to the sample prior to sonication. Solubilized proteins were subject to disulphide reduction and sulphhydral alkylation (200 mM DTT and 200 mM iodoacetamide). Digestion was carried out overnight at 37°C using 20 μg of trypsin (Promega). The digested samples were cleaned using a cation exchange Cartridge Kit for cICAT (Applied Biosystems).

(b) Strong cation exchange chromatography

Samples were brought up to 2 ml with 10 mM KPO₄ (pH = 2.7), 25% ACN buffer and injected onto a Poly sulphoethyl A strong cation exchange chromatography (SCX) column (Poly LC, Columbia, MD). The flow rate was set to 0.5 ml min⁻¹. After equilibration, a 0–35% gradient of 10 mM KH₂PO₄, 25% ACN, 0.5 M KCl was applied for 30 min. Each collected SCX fraction was reduced and transferred to autosampler vials (Dionex/LC Packings, Amsterdam).

(c) One-dimensional reversed-phase chromatography with online mass spectrometry

Liquid chromatography-mass spectrometry/mass spectrometry (LC-MS/MS) analyses of the SCX seminal fluid protein fractions were performed in a Hybrid Quadruple-TOF LC–MS/MS mass spectrometer (QStar Pulsar I, MDS Sciex), equipped with a nanoelectrospray ionization source (Proxeon, Odense, Denmark) fitted with a 10 μm fused-silica emitter tip (New Objective, Woburn, MA). Chromatographic separation was achieved on C18AQ Nano LC and a Zorbax C18 guard column (Agilent Technologies). The mobile phase consisted of 98 : 2 (v/v) water/acetonitrile with 0.1% formic acid. Data were acquired automatically using Analyst QS v. 1.1 software service pack (ABI MDS SCIEX, Concord, Canada). Curtain gas was set at 23, nitrogen was used as the collision gas, and the ionization tip voltage was 2700 V.

(d) Mass spectrometry data analyses

Analyst v. 1.1 software was used to view the information dependent acquisition file, and a built-in Mascot script (1.6b16 ABI—Matrix Science Limited) was used to create the peak lists. Spectra with less than 10 peaks were discarded. MS/MS data were centroided but not de-isotoped. Data were analysed using Mascot v. 2.0 (Matrix Science Limited). Carbamidomethyl cysteine was used as a fixed modification, oxidation of methionine was selected as a variable modification, and up to one missed cleavage was allowed. Spectrometry data were searched against deduced partial proteomes of G. firmus and G. pennsylvanicus accessory glands. The combined database contained 1435 putative protein entries with 32 048 residues. Details of the development of the EST database supporting partial proteomes have been described elsewhere (Andrés et al. 2006).

(e) Genetic analyses: sampling

(i) Populations

We sampled two allopatric populations of each of the cricket species (G. firmus: Guilford, CT and Essex, MD;
(ii) Genes
In total, we analysed polymorphism and divergence data for the entire coding regions of six genes, all of which are expressed in cricket accessory glands: peptidyl-prolyl isomerase 1 (PPlase), guanylate kinase-1 (GuKc), ubiquinone biosynthesis protein COQ7-1 (COQ7) and three seminal proteins (AG-0005F, AG-0308F and AG-0334P). PPlase is an enzyme that accelerates protein folding by catalysing the cis-trans isomerization of proline peptide bonds; GuKc catalyses the ATP-dependent phosphorylation of GMP into GDP; and COQ7 is a central metabolic regulatory protein. These proteins, which do not show male-biased expression, have not been identified in the spermatophore and probably do not have a function in cricket reproduction. The genes that encode these proteins are therefore considered as ‘control genes’. Of the three seminal proteins AG-0308F is a serine protease, whereas the other two seminal proteins are biochemically uncharacterized.

Total RNA from accessory glands of individual crickets was extracted using Trizol Reagent (Invitrogen), reverse transcribed and PCR amplified using gene-specific primers (Andrés et al. 2006; see the electronic supplementary material S1). The PCR products were sequenced using an ABI 3100 automatic sequencer (Applied Biosystems, Foster City, CA). Initial visual identification of single nucleotide polymorphisms were confirmed using TracEdiff and HetScan of the Staden package (Staden 1996). Individual alleles were reconstructed using the Phase algorithm (Stephens et al. 2001). Sequences have been deposited in GenBank (EU669672–EU669818).

(f) Genealogical analyses
For each dataset, an optimal substitution model was determined by hierarchical likelihood ratio tests using Modeltest v. 3.7 (Posada & Crandall 1998; see the electronic supplementary material S2), and the evolutionary relationship among alleles was estimated by maximum-likelihood (ML) methods using PAUP v. 4.0b10 (Swofford 2003). Nodal support was based on 1000 heuristic non-parametric bootstrap replicates. To test the agreement among loci, we used a partition of variance test with 1000 heuristic replicates. To test if G. firmus and G. pennsylvanicus constitute exclusive (monophyletic) groups, we performed exclusivity tests by comparing the increment in the posterior log-likelihood (Δln L) distribution between unconstrained gene genealogies and genealogies constrained to represent the two species as (monophyletic) groups, we performed exclusivity tests by bootstrap replicates. To assess the agreement among loci, we used a partition of variance test with 1000 heuristic replicates. To test if G. firmus and G. pennsylvanicus constitute exclusive (monophyletic) groups, we performed exclusivity tests by comparing the increment in the posterior log-likelihood (Δln L) distribution between unconstrained gene genealogies and genealogies constrained to represent the two species as (monophyletic) groups, we performed exclusivity tests by bootstrap replicates. To assess the agreement among loci, we used a partition of variance test with 1000 heuristic replicates. To test if G. firmus and G. pennsylvanicus constitute exclusive (monophyletic) groups, we performed exclusivity tests by comparing the increment in the posterior log-likelihood (Δln L) distribution between unconstrained gene genealogies and genealogies constrained to represent the two species as (monophyletic) groups, we performed exclusivity tests by bootstrap replicates. To assess the agreement among loci, we used a partition of variance test with 1000 heuristic replicates.

(g) Test of selection
The relative rate of fixation of non-synonymous (dN) and synonymous (dS) substitutions provides an estimate of the selection pressures acting on a given protein. For any set of amino acid residues, when dN/dS = ω = 1, a neutral model of evolution cannot be rejected, whereas ω<1 indicates purifying selection and ω>1 indicates positive selection.

Although the selection parameter ω is commonly calculated using phylogenetic likelihood methods (Goldman & Yang 1994), these methods are unreliable in the presence of intragenic recombination because this process leads to not one, but multiple evolutionary trees along the gene sequence (Anisimova et al. 2003; Wilson & McVean 2006). In this paper we used the method recently developed by Wilson & McVean (2006) to calculate ω in the presence of recombination. This method relaxes the assumption of a single common history for all codons, and performs Bayesian inferences of ω using a population genetics approximation to the coalescent with recombination (Hudson 1983; Li & Stephens 2003). One disadvantage of this method is that it does not provide estimates of dS and dS.

Using OMEGAMAP (Wilson & McVean 2006) we estimated the selection parameter (ω), recombination rate (ϕ), transition–transversion ratio (κ) and the rate of synonymous transversion μ for each gene. We used improper inverse prior distributions for all parameters with means ω = 1, ϕ = 0.07, ς = 3.6 and μ = 0.3. Both ω and ϕ were modelled as constants (i.e. all sites are assumed to share common values). The frequency of codons was assumed to be equal, and the number of alignment orderings was set to 10. The number of iterations of the Markov chain Monte Carlo (MCMC) algorithm was 250,000 with a burn-in of 25,000 and a thinning of 1000. For each gene, two independent convergent runs were merged to provide the posterior distributions of the estimated parameters. The effective sample size for the estimated parameters was always more than 100.
Table 1. Field cricket seminal proteins identified by shotgun proteomics (two-dimensional LC/LC–MS/MS). (Gene expression pattern is as described in Andrés et al. (2006); M, male-biased expression; M/F, expressed in both sexes). All proteins have an associated probability of being correctly identified above 99.9% (based on Mascot MudPIT scores). NP is the number of digested peptides that significantly matched each protein. SC represents the percentage of protein sequence identified in the sample. Only peptide hits of eight amino acids or longer and showing a 99% probability of being correct were accepted for protein identification. In the case of single-peptide hits, the probability threshold was increased to 99.9%.

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suggesting that the MCMC chains were run long enough to obtain accurate estimates of the parameters.

Statistical differences in the selection parameter (\( \omega \)) among loci were inferred using Bayesian pairwise comparisons. If the two given loci A and B are under the same evolutionary constraints, the difference between the posterior distribution of \( \omega \) estimated from each locus should not be distinguishable from zero. Thus, the 95% credible interval of the distribution of \( \omega_A - \omega_B \) should include zero. Otherwise, these two \( \omega \) values are significantly different.

3. RESULTS

(a) Proteomic analyses

We combined data from two accessory gland EST libraries (Andrés et al. 2006) with shotgun (LC/LC–MS/MS) peptide sequencing of proteins extracted from field cricket spermatophores (table 1). This dataset represents a pool of two mass spectrometry analytical runs on two independently extracted protein samples (n=6 males/sample). Comparing the spectrometry data of both runs against our Gryllus accessory gland proteome database (n=1435 putative proteins), we identified 30 genes that are expressed in the male accessory gland. Of these genes, 22 appear to encode proteins that are components of the spermatophore coat or the seminal fluid (table 1); the other eight are probably associated with residual sperm in our sample (see the electronic supplementary material S3). Most of the identified genes are expressed exclusively or more strongly in males and encode proteins of unknown function, i.e. proteins that lack any functional homology or structural similarity to biochemically characterized proteins (table 1). Among these ‘unknown’ proteins are six (encoded by AG-0001F, AG-0076F, AG-0312F, AG-0313F, AG-0315F and AG-0317F) that have a highly repetitive primary structure of small amino acid residues (Ala, Leu) and a predicted helical structure. These are probably proteins of the spermatophore coat (Andrés et al. 2006). Three loci (AG-0159F, AG-0308F and AG-0508F) encode proteins with predicted serine protease activity; proteases are known to be an important component of the seminal fluid in other insects (e.g. Mueller et al. 2005). One locus codes for a presumed nucleotidase and the remaining 12 genes encode seminal fluid proteins for which structures/functions have not been identified. Moreover, by expanding our search to include general peptide databases (MSDB), we have also identified additional ‘housekeeping proteins’ (see the electronic supplementary material S3). Many of these proteins (e.g. \( \beta \)-actin, \( \gamma \)-actin, \( \alpha \)-tubulin, ATP-synthetase) are probably components of sperm.

(b) Genealogical analyses

In order to contrast genealogical patterns in seminal proteins with those for control genes, we examined patterns of molecular variation for the entire coding regions of six genes in allopatric populations of G. firmus and G. pennsylvanicus. The three control genes (COQ7, GuKc and PPase) are expressed in accessory gland, but have not been identified as components of the seminal fluid and are not probably involved in reproductive functions. The three putative seminal protein genes were chosen from the list in table 1 and include a serine protease (AG-0308F) and two genes encoding proteins of unknown function (AG-0005F and AG-0334P). Sequence datasets ranged from 432 bp for PPase to 957 bp for COQ7. Excluding autapomorphies, PPase, COQ7 and GuKc contained very few segregating sites (PPase, 0/432; COQ7, 1/568; GuKc 5/997), whereas two of the seminal protein genes have a higher number of phylogenetically informative sites (24/924 and 9/958 for AG-0005F and AG-0334P, respectively).

For all variable genes, inferred genealogies showed an incongruent phylogenetic signal (partition homogeneity test, \( p < 0.01 \)). For each locus, we used a Bayesian framework to test the hypothesis that G. firmus and G. pennsylvanicus form exclusive genealogical groups (see §2). The hypothesis of exclusivity was rejected for variable control genes; COQ7 and GuKc did not show evidence of species-specific clustering (and PPase exhibited no variation). For COQ7 and GuKc, the credible interval of the difference between the posterior distribution of the likelihood scores of unconstrained and constrained (i.e. reciprocally monophyletic) genealogies is entirely positive (does not overlap 0; table 2). Thus, there is over 5% chance that the true value of the difference between unconstrained and monophyletic genealogies is 0.

In addition to examining patterns of variation for three accessory gland expressed control genes, we also reanalysed data from Broughton & Harrison (2003) on patterns of variation in G. firmus and G. pennsylvanicus for four
nuclear gene introns (table 2). Data from these control genes (EF-1α, calmodulin, phosphoglucone isomerase and cytochrome c) reveal extensive haplotype sharing and also fail to support an hypothesis of exclusivity for the two cricket species.

In contrast to the results for control genes, two of the seminal protein genes (AG-0005F and AG-0334P) exhibit nearly fixed differences between the cricket species, and the gene genealogies of these two loci do not differ significantly from one showing an exclusive relationship between the two species (table 2). The ML tree for AG-0334P (figure 1) reveals two major, well-supported clades. One clade includes all haplotypes from G. firmus populations plus one haplotype from a single G. pennsylvanicus population. The second clade includes all but three haplotypes from G. pennsylvanicus population. The ML tree for AG-0005F also shows significant divergence between the two species, with only haplotypes from G. firmus in one well-supported clade, and all haplotypes from G. pennsylvanicus populations, together with three haplotypes from G. firmus populations, in the second clade (figure 1). Correspondingly, the estimated levels of molecular divergence between species for AG-0334P and AG-0005F are higher than for those genes that show non-exclusive gene genealogies (table 3). Nucleotide sequence data for the third seminal protein gene, AG-0308F, showed a pattern of molecular variation similar to that for control genes, with a very small proportion of informative sites (3/666) and no evidence of exclusivity.

For none of the loci were the results significantly affected by the type of phylogenetic analysis applied. For all loci, ML and maximum parsimony trees had consistent topologies (Wilcoxon signed-rank test p values ranged from 0.629 to 1). Distance-based analyses (which group alleles on the basis of overall similarity reflecting approximate patterns of relatedness among alleles) and parsimony networks (reflecting uncertainty generated by recombination) also yielded the same results as the ML gene genealogies.

Comparisons of amino acid sequences within AG-0005F or AG-0334P revealed substantial differences between alleles’ characteristic of the two species. With the exception of a single allele found in the Scranton, PA population, AG-0334P is unambiguously sorted by species. The clades supporting ‘exclusivity’ of G. firmus and G. pennsylvanicus have high bootstrap support values (71 and 91%, respectively). AG-0334P has two principal allelic forms that are characterized by three radical and three conservative amino acid differences (Asn75Thr, His85Leu, Ile166Asn, Gly168Cys, Asp191Glu and Glu205Lys; see the electronic supplementary material S4). The ML genealogy for AG-0005F also shows two clades (94% bootstrap support) associated with two species characteristic allelic forms of the protein. These alleles are highly divergent; aside from the G. pennsylvanicus alleles found in presumed G. firmus populations, there are five fixed radical amino acid substitutions between the two species (Phe117Val, Ile167Thr, Lys182Gln, Gln228Leu(His) and Leu280Phe; see the electronic supplementary material S5). The topologies are robust to model choice, and different amino acid substitution models (JTT, PAM or PMB) yielded the same ML topologies.

(c) Test of selection

Seminal protein loci that show substantial differentiation between G. firmus and G. pennsylvanicus (AG-0005F and AG-0334P) exhibit an elevated rate of non-synonymous substitutions (dKs) compared with loci that fail to show evidence of species-specific clustering (AG-0308F, COQ7 and GuKc). The 95% credible intervals of the posterior probability distribution of the selection parameter (ω) for the pair of genes showing strong haplotype frequency differences between species include values greater than 1, as expected if positive selection plays an important role in the diversification of these genes (table 4). Pairwise comparisons show that the ω values for these genes are generally higher than for genes showing no evidence of differentiation between species (table 5).

4. DISCUSSION

During the speciation process, genomes of diverging lineages will be mosaics with respect to molecular genealogy (Ting et al. 2000). Under a neutral model of divergence, stochastic lineage sorting that leads to exclusivity or reciprocal monophyly is expected to occur over long periods of time in organisms such as field crickets with large effective population sizes and only a single generation each year. Throughout most of the genome, diverging lineages may continue to share alleles or haplotypes owing to introgression and/or the retention of ancestral polymorphisms. By contrast, the genealogies of speciation/barrier genes and genes responsible for lineage-specific adaptations are more likely to reveal differentiation or exclusivity between species.

Many recent studies of closely related species (flies, mice, sunflowers, moths and butterflies) have demonstrated genealogical discordance and/or differential
These studies, which represent genome scans or scans of the sex chromosome, have also defined genome regions that show little evidence of introgression or for which closely related species are exclusive groups (Machado & Hey 2003; Panithanarak et al. 2004; Dopman et al. 2005). One pattern that has emerged is that genome regions associated with fixed differences in chromosome arrangements introgress less (Rieseberg et al. 1999; Noor et al. 2001; Ortiz-Barrientos et al. 2002) and show accelerated rates of protein evolution (Navarro & Barton 2003). In the absence of a linkage map for field crickets, we cannot examine patterns of differentiation at defined genomic regions. Instead, we have used a proteomic approach to target a particular class of genes (seminal proteins), in a search for regions of exclusivity or differentiation within the cricket genome.

Because the cricket species G. pennsylvanicus and G. firmus have diverged very recently, we expect that most regions of the genome will continue to reveal shared introgression (Rieseberg et al. 1999; Beltran et al. 2002; Machado & Hey 2003; Ting et al. 2000; Panithanarak et al. 2004; Dopman et al. 2005; Payseur & Nachman 2005). These studies, which represent genome scans or scans of

ancestral polymorphisms (Broughton & Harrison 2003). Indeed, previous efforts to identify diagnostic differences between the two species have generally been unsuccessful and most genomic regions that have been surveyed exhibited evidence of shared polymorphisms (Harrison & Bogdanowicz 1997; Broughton & Harrison 2003).

Because field cricket females are highly promiscuous, and because fertilization barriers are partly responsible for reproductive isolation between G. firmus and G. pennsylvanicus (Harrison 1983), genes encoding seminal proteins are likely to be under strong diversifying selection and may be involved in barriers to gene exchange. If so, genes encoding seminal proteins should be strongly differentiated, and their genealogies should be congruent with the reproductive differences between these two species.

The gene genealogies of the ‘control loci’ examined in this study (COQ7, GuKc and PPase) confirmed what we learned from previous genealogical analyses of four nuclear gene introns (Broughton & Harrison 2003). Although levels of variation differ substantially among these loci, G. firmus and G. pennsylvanicus do not form exclusive groups at any locus and much of the polymorphism appears to predate lineage splitting (i.e. shared alleles do not show geographical structure). Thus, as expected, a large portion of the field cricket genome has remained undifferentiated after the evolution of reproductive isolation. By contrast, two of the three loci that encode seminal proteins (AG-0005F and AG-0334P) are highly divergent between species, and sequence differences at these loci include radical amino acid substitutions that could have functional significance for the reproductive biology of the crickets. Given the recent divergence of the two cricket species (estimated to be 0.1N_e by Broughton & Harrison 2003) random sorting of ancestral polymorphisms is improbable. Thus, the observed pattern suggests that AG-0005F and AG-0334P might play an important role in species differentiation. However, the extent to which these two genes represent outliers in the cricket genome needs to be confirmed by a more extensive survey of both ‘control’ and seminal protein genes.

During the early stages of the speciation process, positive selection is likely to be responsible for the origin of genomic differentiation between diverging lineages. Lineage-specific selective sweeps will produce regions of exclusivity surrounding loci under selection. Our results suggest that the pattern of differentiation observed for AG-0005F and AG-0334P may be a consequence of rapid evolution due to selection acting on these loci. The selection coefficient parameter (ω) is significantly higher at these loci than those loci showing no evidence of selection. Positive selection is likely to be responsible for the observed pattern of expression (Duret & Mouchiroud 2000; Winter et al. 2004). The ability of our tests to detect positive selection is relatively limited (see Hughes 2007), and the estimated distribution of ω values cannot unequivocally rule out a scenario of lack of constraint.

Table 4. Detection of selection in the presence of intragenic recombination. (Values represent the posterior probability means of the transition–transversion ratio (κ), the rate of synonymous transversion (μ), recombination rate (ρ) and selection parameter (ω). ω values in parentheses represent the 95.5% credible intervals of the posterior probability distributions. Values in italics represent ω values with credible intervals that include estimates consistent with positive selection acting on that locus (i.e. ω > 1.).

<table>
<thead>
<tr>
<th>locus</th>
<th>κ</th>
<th>μ</th>
<th>ρ</th>
<th>ω</th>
</tr>
</thead>
<tbody>
<tr>
<td>AG-0005F</td>
<td>2.465</td>
<td>0.166</td>
<td>0.075</td>
<td>0.602 (0.342–1.173)</td>
</tr>
<tr>
<td>AG-0308F</td>
<td>4.541</td>
<td>0.054</td>
<td>11.576</td>
<td>0.043 (0.010–0.123)</td>
</tr>
<tr>
<td>AG-0334P</td>
<td>0.858</td>
<td>0.055</td>
<td>0.017</td>
<td>1.091 (0.201–2.702)</td>
</tr>
<tr>
<td>COQ7</td>
<td>10.917</td>
<td>0.059</td>
<td>10.78</td>
<td>0.216 (0.017–0.613)</td>
</tr>
<tr>
<td>GuKc</td>
<td>2.462</td>
<td>0.072</td>
<td>0.018</td>
<td>0.325 (0.037–0.736)</td>
</tr>
</tbody>
</table>

Table 5. Posterior mean of the difference between the estimated ω values for each pair of loci (e.g. AG-0005F–AG-0308F) and its 95% credible interval. (Two ω values are significantly different if the credible interval of their difference does not include zero *p < 0.05, **p < 0.01.)

<table>
<thead>
<tr>
<th></th>
<th>AG-0005F</th>
<th>AG-0308F</th>
<th>AG-0334P</th>
<th>COQ7</th>
<th>GuKc</th>
</tr>
</thead>
<tbody>
<tr>
<td>AG-0005F</td>
<td>0.460**</td>
<td>-0.591</td>
<td>0.369*</td>
<td>0.203</td>
<td></td>
</tr>
<tr>
<td>AG-0308F</td>
<td>(0.197–0.975)</td>
<td>(-2.273 to 0.296)</td>
<td>(0.132–1.417)</td>
<td>(-0.512 to 0.738)</td>
<td></td>
</tr>
<tr>
<td>AG-0334P</td>
<td>-1.071**</td>
<td>(-3.754 to 0.179)</td>
<td>-0.214</td>
<td>(-0.992 to 0.014)</td>
<td></td>
</tr>
<tr>
<td>COQ7</td>
<td>0.895**</td>
<td>(0.321–1.973)</td>
<td>0.790*</td>
<td>(0.043–3.500)</td>
<td></td>
</tr>
</tbody>
</table>

For both AG-0005F and AG-0334P alleles characteristic of one species are found at low frequency in the other species. For AG-0005F, two crickets from Guilford and one cricket from Essex each carry one G. pennsylvaniaicus allele; for AG-0334P, a single Scranton individual carries a G. firmus allele. This pattern could be explained either by persistence of shared ancestral polymorphisms or by recent hybridization and introgression. The similarity of the ‘foreign’ alleles in one species to those currently found in the other species argues in favour of recent introgression. The consequences of the presence of foreign alleles at AG-0005F and AG-0334P for the reproductive phenotype of the individual crickets are not yet clear. That the two species share alleles, however, does not preclude a role for the protein products of AG-0005F and AG-0334P in barriers to gene exchange. Given our allele sampling, more extensive population genetic analyses are necessary to assess the relative extent of allele sharing at these loci. Furthermore, the effects of variation in amino acid sequence can be assessed by testing the reproductive phenotype of knock-down (RNAi) males as well as by pairing males and females with different allelic combinations.

A proteomics approach for identifying candidate speciation/barrier genes certainly shows great promise. Using a combination of genomic, proteomic, phylogenetic and population genetic techniques we have been able to identify a combination of genomic, proteomic, phylogenetic and population genetic techniques we have been able to identify candidate barrier genes in field crickets, but functional analyses are obviously required before we can understand the role of seminal proteins in reproductive isolation.

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