Better than fish on land? Hearing across metamorphosis in salamanders

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Early tetrapods faced an auditory challenge from the impedance mismatch between air and tissue in the transition from aquatic to terrestrial lifestyles during the Early Carboniferous (350 Ma). Consequently, tetrapods may have been deaf to airborne sounds for up to 100 Myr until tympanic middle ears evolved during the Triassic. The middle ear morphology of recent urodeles is similar to that of early ‘lepospondyl’ microsaur tetrapods, and experimental studies on their hearing capabilities are therefore useful to understand the evolutionary and functional drivers behind the shift from aquatic to aerial hearing in early tetrapods. Here, we combine imaging techniques with neurophysiological measurements to resolve how the change from aquatic larvae to terrestrial adult affects the ear morphology and sensory capabilities of salamanders. We show that air-induced pressure detection enhances underwater hearing sensitivity of salamanders at frequencies above 120 Hz, and that both terrestrial adults and fully aquatic juvenile salamanders can detect airborne sound. Collectively, these findings suggest that early atympanic tetrapods may have been pre-equipped to aerial hearing and are able to hear airborne sound better than fish on land. When selected for, this rudimentary hearing could have led to the evolution of tympanic middle ears.

1. Introduction

Sound detection and auditory perception are used to navigate, communicate, find food and avoid predators [1]. Hearing hence provides vital information about the surrounding environment to a large range of both aquatic and terrestrial animals. Auditory systems of vertebrates have been shaped by evolution to cope with the physical properties of the two very different media of air and water. Although the functional unit of hearing in all vertebrates is the hair cell, sensitive to displacement [2,3], various transduction elements have evolved to enable detection of the particle motion and pressure components of sound.

Because the impedance of animal tissue is close to the impedance of water, sound waves can travel almost unhindered to the hair cells of the inner ear maculae in aquatic vertebrates. In fish auditory systems, otolithic organs enable hair cell deflection by differential inertial movements of otolith and hair cells [4,5], and so the adequate stimulus for these sensory organs is particle motion in the form of acceleration [6]. To evolve pressure-sensitive ears, pressure waves need to be converted to detectable particle motion [7,8]. In many aquatic vertebrates, this is accomplished by gas-filled structures such as swim bladders [7,9] or bullae [10], or secondarily by middle ear cavities [11]. The enclosed air in such cavities increases the available particle motion when ensonified by a pressure wave providing up to two orders of magnitude more acceleration than particle motion in the surrounding water [12].

In contrast to aquatic vertebrates, tetrapods in air have the problem that the impedance of tissue is much higher than the impedance of air, and thus most of the sound energy is reflected on the air–tissue boundary. Hence, early tetrapods faced an impedance problem as they moved from aquatic to terrestrial lifestyles.
during the Early Carboniferous. The solution to this problem in recent tetrapods is the evolution of the tympanic middle ear, which converts sound pressure in air to particle motion in the fluid of the inner ear. According to the recent palaeontological record, however, the tympanic middle ear did not appear until the Early Triassic, where tympanic middle ears evolved independently in all the tetrapod lineages [13]. Presumably, terrestrial vertebrates were therefore not adapted to detect aerial sound pressure for up to 100 Myr, raising the question of whether early tetrapods were functionally deaf.

The impedance mismatch is also faced by some extant amphibious vertebrates. In most anurans, it is overcome by development of a tympanic middle ear during or after metamorphosis [14], resulting in improved pressure sensitivity of the adults in addition to the particle-motion-sensitive ear found in the aquatic juveniles [15]. However, not all amphibians develop a tympanic middle ear during metamorphosis. For example, none of the urodèles have tympanic middle ears [16]; instead, the columella articulates distally with the squamosal or palatoquadrate of these animals. Additionally, the urodele middle ear also contains the operculum, which is connected to the scapula of the shoulder girdle through the opercularis muscle and has been proposed to aid the transmission of substrate vibrations into the inner ear via the forelegs [17], play a role in airborne hearing by bone conduction [18] or function as a protective mechanism against loud sound exposures [19]. The morphology of the urodele auditory system resembles that of early 'lepospondyl' microsaurs tetrapods in the shape of the columella and the lack of a tympanic middle ear [13,20]. Equally important, it can be regarded as a potential model for the intermediate evolutionary developmental stage between the aquatic adapted auditory systems of tetrapod ancestors (exemplified by the auditory system in recent lungfish) and the auditory systems of recent tympanic tetrapods adapted to aerial hearing seen in most anurans (figure 1). Experimental studies on the hearing capabilities in recent urodèles are therefore instructive for uncovering the evolutionary and functional drivers behind the shift from aquatic to aerial hearing in early tetrapods.

However, only a few experimental studies have investigated hearing and vibration detection of urodèles, and changes across metamorphosis have never (to our knowledge) been studied. Overall, the morphology of the urodele middle ear implies good sensitivity to substrate vibrations, but poor sensitivity to aerial sound, suggesting that urodèles may be no more adapted to aerial hearing than are fish. In agreement with the morphological expectations, urodèles have previously been shown to be very sensitive to substrate vibrations [19,21–25], but surprisingly, earlier studies also suggest that urodèles are able to detect airborne sound [19]. Urodèles would seem unable to use pressure-to-particle motion transduction by air volumes in their lungs to enable underwater pressure detection as there is no mechanical connection between the lungs and the inner ears. Yet an earlier study indicates that urodèles may detect underwater sound pressure using an air volume in the mouth cavity for pressure-to-particle motion transduction [26].

Here, we combine imaging techniques with neurophysiological measurements in both water and air in an attempt to resolve how the change from aquatic larva to terrestrial adult through metamorphosis affects the morphology of the ears and the sensory capabilities of urodèles. To allow for a controlled and thus uniform metamorphosis, the axolotl (Ambystoma mexicanum) was chosen as our experimental animal. This neotenic salamander is a convenient choice for comparative studies across metamorphosis as this can easily be induced by thyroid hormone treatment and thereby controlled in an experimental study. Additionally, we investigate the vibration detection and aerial hearing of adult specimens of the closely related tiger salamanders (Ambystoma tigrinum) to compare hearing abilities of aquatic and terrestrial salamanders. Our results show that both juvenile and adult salamanders are able to detect airborne sound. Furthermore, pressure detection is found to enhance underwater hearing sensitivity of salamanders at frequencies above 120 Hz. In combination, these findings suggest that early atypic tympanic tetrapods may have been pre-equipped to aerial hearing and able to hear airborne sound better than fish on land.

2. Material and methods

The study was conducted using 20 axolotls (mass: 68.5 ± 20.3 g and total length: 21.5 ± 2.8 cm, mean ± s.d.) and six adult tiger salamanders (mass: 38.0 ± 4.5 g and total length: 20.7 ± 1.5 cm). Both axolotls and tiger salamanders were obtained commercially and kept in a 12 L:12 D cycle at room temperature (approx. 20 °C). The salamanders were anaesthetized before measurements by submergence in a 0.25% Benzocaine (Sigma-Aldrich, St Louis, MO, USA) water solution until they failed to execute the righting reflex. Benzocaine was further added to the water, resulting in a 0.05% solution to uphold the anaesthesia during underwater measurements. The juvenile axolotls were kept moist during measurements in air by wrapping them in wet paper towels and dripping them with the Benzocaine solution several times. Animals recovered from anaesthesia in about 20 min when returned to benzocaine-free water after measurements. The animals were sacrificed at the end of the sensitivity experiments by submergence in a high-concentration Benzocaine solution and fixed in buffered 4% formaldehyde solution (VWR, Leuven, Belgium) for later computed tomography (CT) scanning. The fixation did not have any significant effect on the preparations.

(a) Metamorphosis

The vibration and hearing sensitivity was determined in the 20 juvenile axolotls. Next, metamorphosis was induced in 12 axolotls by addition of thyroxin hormone (T4; Sigma-Aldrich) to the water of their aquarium [27]. The remaining eight
juveniles were used as controls kept under the same conditions in identical aquaria to enable comparison with animals that also got older, but did not metamorphose. Thirty days after reaching stage 4 of the metamorphosis [28], the adult axolotls, along with the eight controls, were then re-tested to investigate changes across metamorphosis.

(b) Experimental set-up and calibration

The neurophysiological experiments in air were conducted in a combined acceleration and sound pressure set-up [29]. The salamanders had their head resting on a shaker platform 80 cm below a loudspeaker to determine acceleration and sound pressure threshold, respectively. The shaker (Bruel & Kjær Vibration Exciter, Type 4809, Naerum, Denmark) was calibrated using a Bruel & Kjaer Accelerometer (Type 4381) calibrated using a Bruel & Kjaer Calibration Exciter (Type 4294) with an output of 10 ms⁻² at 159.15 Hz. The speaker (8 inch V8 installation speaker, Tannoy Ltd, Coatbridge, UK) was calibrated using a 4 inch free field microphone (Type 40AF, GRAS, Holte, Denmark), calibrated with a Bruel & Kjaer Acoustical Calibrator (Type 4231, Bruel & Kjaer) with an output of 94 dBrms re 20 Pa at 1000 Hz.

The underwater experiments were conducted in a standing wave tube set-up [30] where an underwater loudspeaker was placed in the bottom of a 2 m long and 30 cm diameter water-filled steel tube with 1 cm thick walls. Sound stimulation created standing waves in the tube and hence the underwater hearing sensitivity could be investigated under different particle motion-to-pressure conditions by changing the measuring depth in the tube. Both pressure and particle motion of the sound field in the tube was calibrated using two hydrophones (Reson TC 4013), with a flat frequency response in the frequency range used. The hydrophones were calibrated using a Bruel & Kjaer hydrophone calibrator (Type 4223) with an output of 94 dBa₂s re 20 µPa at 1000 Hz. The underwater experiments were conducted in a standing wave tube and hence the underwater hearing sensitivity could be investigated under different particle motion-to-pressure conditions by changing the measuring depth in the tube. Both pressure and particle motion of the sound field in the tube was calibrated using two hydrophones (Reson TC 4013), with a flat frequency response in the frequency range used. The hydrophones were calibrated using a Bruel & Kjaer hydrophone calibrator (Type 4223) with an output of 94 dBa₂s re 20 µPa at 1000 Hz.

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information of cephalic and thoracic anatomy of the salamanders under study: a Scanco Medical XtremeCT system and a Scanco Medical μCT-35 system (Brüttisellen, Switzerland). A staining protocol was applied to some of the specimens to reveal soft tissue structures [34]. After formaldehyde fixation, samples were washed for three weeks in phosphate buffer to remove any residual formaldehyde, thereafter immersed for four weeks in diluted Lugol’s solution (0.33% I2 and 0.67% KI). Imaging parameters of the XtremeCT system were: 63.63 × 28.54 mm² field of view; 776 × 348 matrix; 0.082 mm slice thickness; 59.4 kVp tube voltage; 119 μAs tube current, resulting in an acquisition time of 35 min. Higher-resolution data were acquired using the μCT-35 system with parameters: 21.04 × 23.56 mm² field-of-view; 1403 × 1571 matrix; 0.015 mm slice thickness; 55 kVp tube voltage; 116 μA tube current, resulting in an acquisition time of 10 h.

The XtremeCT system was used on the anterior of three stained and three unstained juvenile axolotls, three stained and three unstained adult axolotls, and two unstained tiger salamanders. In addition to these specimens, a single tiger salamander was stained, and of these specimens one stained and one unstained specimen from each phenotype were selected for high-resolution μCT. Images of interrelated samples were registered and three-dimensional models generated using AMIRA v. 5.6 as described by Ruthensteiner & Heß [35].

3. Results

(a) Morphology

Metamorphosis caused distinct and easily recognizable changes in the outer morphology of the axolotls. The animals showed an average weight reduction (± s.d.) of 38 ± 8%, and tail keel and gills shrunk, and were completely lost in the adult stage. X-ray CT data showed that the columella is free in both the juvenile and adult axolotl, but fused to the otic capsule in the tiger salamander (figure 2; also see interactive three-dimensional models in electronic supplementary material, figures S4–S6). The opercularis muscle connecting scapula and operculum was easily recognized in iodine-stained specimens of tiger salamanders and adult axolotls, but could not be found in the juvenile axolotls. The cartilaginous operculum could not be recognized in the CT scans in any specimens (figure 2), but a large aperture was found in the otic capsule posterior to the columella (i.e. at the location of the operculum) in both juvenile and adult salamanders. Dissections of inner and middle ears showed that all three phenotypes had a movable operculum, which in the adult axolotls and tiger salamanders clearly responded to movement of the scapula. Moreover, the columella was movable in both juvenile and adult axolotls, but rigid in tiger salamanders, confirming the findings from the CT data.

CT was also used to determine air volumes in lung and mouth cavities of the axolotls (table 1). In awake animals, air volumes were only found in the lung cavities, whereas no air was observed in the mouth cavities. Anaesthesia and handling reduced lung air volumes in both groups, but mostly in the adult axolotls (table 1; paired t-test: juveniles: t = −3.299, p = 0.013; 320 Hz: t = −3.096, p = 0.017; 200 Hz: t = −5.362, p = 0.001; 320 Hz: t = −9.089, p < 0.001; 640 Hz: t = −7.761, p < 0.001). Particle motion thresholds determined under high-particle-motion conditions and pressure thresholds determined under high-pressure conditions are plotted in figure 3b and 3c, respectively. All thresholds found were at least 15 dB above the octave noise level of the frequencies tested. As the critical bands in amphibians are smaller than one octave [36,37], the octave noise level generates an upper bound estimate of the masking by ambient noise.

(b) Adequate stimuli under water

No significant change was found in particle motion thresholds for 80 and 120 Hz from high particle motion to high-pressure conditions for either juvenile or adult axolotls (figure 3a). At frequencies above 120 Hz, however, particle motion sensitivity was significantly increased in both these phenotypes under high-pressure conditions relative to sensitivity found under high-particle-motion conditions (figure 3a; paired t-test: juveniles: 160 Hz: t = −2.868, p = 0.024; 200 Hz: t = −3.299, p = 0.013; 320 Hz: t = −7.906, p < 0.001; 640 Hz: t = −5.099, p = 0.004; adults: 160 Hz: t = −3.096, p = 0.017; 200 Hz: t = −5.362, p = 0.001; 320 Hz: t = −9.089, p < 0.001; 640 Hz: t = −7.761, p < 0.001). Particle motion thresholds determined under high-particle-motion conditions and pressure thresholds determined under high-pressure conditions are plotted in figure 3b and 3c, respectively. All thresholds found were at least 15 dB above the octave noise level of the frequencies tested. As the critical bands in amphibians are smaller than one octave [36,37], the octave noise level generates an upper bound estimate of the masking by ambient noise.

(c) Vibration and sound pressure sensitivity in air

Vibration sensitivity curves (vibrograms) of both axolotls and tiger salamanders had a W-shape with two distinct peaks (figure 3d). Juvenile axolotls had best frequencies of 40 and 160 Hz with mean thresholds (± s.e.m.) of −46 ± 1.1 and −44 ± 1.4 dB re 1 m s⁻², respectively, whereas adult axolotls were less sensitive and had higher best frequencies of 80 and 240 Hz with mean thresholds of −41 ± 2.4 and −27 ± 2.4 dB.
Figure 3. Hearing and vibration sensitivity of juvenile and adult axolotls, and adult tiger salamanders in (a–c) water and (d–g) air. (a) Relative change in particle motion thresholds from high-particle-motion to high-pressure depth for juvenile (blue squares) and adult axolotls (red triangles) along with the change in particle motion-to-pressure ratio (black line). Asterisks indicate statistical significance (paired t-test, *p < 0.05; **p < 0.01). (b) Average particle motion audiograms of juvenile (blue squares) and adult axolotls (red triangles) from the high-particle-motion depth. (c) Average pressure audiogram of juvenile (blue squares) and adult axolotls (red triangles) from the high-pressure depth. (d) Vibration sensitivity of juvenile axolotls (blue squares), adult axolotls (red triangles) and tiger salamanders (green diamonds) in response to vertical shaker vibrations. (e) Sound pressure sensitivity of juvenile axolotls (blue squares), adult axolotls (red triangles) and tiger salamanders (green diamonds). (f) Average change in vibration thresholds of axolotls across the metamorphosis (red triangles) along with juvenile controls (blue squares). Asterisks indicate statistical significance (paired t-test, *p < 0.05; **p < 0.01). (g) Average change in sound pressure thresholds of axolotls across the metamorphosis (red triangles) along with juvenile controls (blue squares). Asterisks indicate statistical significance (paired t-test, *p < 0.05; **p < 0.01). Bars are in all plots ± s.e.m. n-values are indicated in each plot.
re 1 m s$^{-2}$. Adult tiger salamanders had a lower best frequency of 40 Hz with a mean threshold of $-45 \pm 1.4$ dB re 1 m s$^{-2}$ and a high best frequency of 240 Hz with a mean threshold of $-42 \pm 2.7$ dB re 1 m s$^{-2}$. Individual thresholds were at least 16 dB above the octave noise level of the frequencies tested.

Sound pressure sensitivity curves were also W-shaped (figure 3c). All groups had best frequencies of 80 and 320 Hz. Juvenile axolotls had mean thresholds of $77 \pm 1.5$ and $82 \pm 1.1$ dB re 20 $\mu$Pa, adult axolotls $81 \pm 1.2$ and $79 \pm 1.6$ dB re 20 $\mu$Pa, and tiger salamanders had mean thresholds of $78 \pm 2$ and $83 \pm 1.6$ dB re 20 $\mu$Pa, respectively. All thresholds were at least 29.5 dB above the octave noise level of the frequencies tested. Further, sound-induced shaker vibrations were below vibration thresholds at all frequencies tested for all three phenotypes.

(d) Change across metamorphosis

Vibration thresholds were increased significantly across metamorphosis at 40, 160, 200, 320 and 640 Hz (figure 3f; paired t-test: 40 Hz: $t = 3.952, p = 0.003; 160$ Hz: $t = 5.385, p < 0.001$; 200 Hz: $t = 4.732, p = 0.001; 240$ Hz: $t = 4.186, p = 0.002$; 320 Hz: $t = 2.738, p = 0.023; 640$ Hz: $t = 3.011, p = 0.015$), whereas no significant difference was found in the control group. Neither the sound pressure thresholds of the metamorphosed axolotls nor of the control group changed significantly across metamorphosis, although thresholds of 160–240 Hz were lower in the adult than in the juvenile stage (figure 3g).

4. Discussion

Here, we used evoked potential measurements to determine the underwater hearing of both juvenile and adult axolotls to test the null hypothesis that urodeles are unable to detect sound pressure under water. We show that particle motion is the adequate stimulus at frequencies up to 120 Hz. Sound pressure is, however, the adequate stimulus at higher frequencies, and we therefore reject the null hypothesis. Evoked potential measurements were also used to determine aerial hearing and vibration sensitivity to elucidate how the auditory abilities of urodeles are affected by the change from an aquatic to a terrestrial lifestyle. Specifically, we wanted to test the null hypothesis that terrestrial adult urodeles, having no special adaptations to aerial hearing, are no better in detecting airborne sound than fully aquatic vertebrates such as fish. It is demonstrated that urodeles indeed are more sensitive to aerial sound than fully aquatic vertebrates, and thus we find that urodeles are better than fish on land when it comes to hearing.

(a) Underwater sound detection

We investigated the underwater hearing capabilities of the axolotl in a standing wave tube system where the particle motion-to-pressure ratio of the sound field changes with depth. Sound detection can therefore be investigated under both high-particle-motion conditions and high-pressure conditions, allowing us to establish the adequate stimulus. Similar hearing sensitivity (in terms of particle motion) under both conditions suggests detection of particle motion, whereas increased sensitivity from high-particle-motion to high-pressure conditions demonstrates pressure detection. No difference was found in particle motion thresholds at frequencies of 80 and 120 Hz for either juvenile or adult axolotls (figure 3o). At frequencies above 120 Hz, however, thresholds determined under high-pressure conditions were significantly lower than thresholds determined under high-particle-motion conditions (figure 3o). Our results therefore show that the adequate stimulus is particle motion at low frequencies (figure 3o), but that both juvenile and adult axolotls are able to detect sound pressure in water at frequencies above 120 Hz (figure 3c).

(b) Detection of substrate vibrations

While sound pressure is the adequate stimulus of ears adapted to aerial hearing, good vibration sensitivity may enable atypical, terrestrial vertebrates to use substrate vibrations as a source of information regarding potential prey, predators and conspecifics, and for communication, as shown for many animals [38].

We found that both axolotls and tiger salamanders are very sensitive to vertical substrate vibrations (figure 3d), consistent with both sensitivity and frequency ranges found in earlier studies [19,21–25]. The high sensitivity is underlined by the fact that thresholds determined by evoked potentials may be 10–30 dB above thresholds determined by single cell recordings or behavioural studies in a variety of animals [39–41]. Moreover, the low-frequency vibration sensitivity found here is comparable with the vibration sensitivity of lungfish [30,42] in terms of best frequency and sensitivity, but whereas the lungfish vibrogram is U-shaped, the salamander vibograms were W-shaped, having an additional peak at higher frequencies. In frogs, both the saccule and the amphibian papilla are involved in detection of substrate vibrations: the saccule being most sensitive at frequencies below 100 Hz, and the amphibian papilla to frequencies between 60 and 600 Hz [43–45]. Coupling of vibrations to the ventral surface cause endolymphatic displacements through opercular vibrations at 50–400 Hz in urodeles [46], and the amphibian papilla is therefore probably also responsible for detection of high-frequency substrate vibrations in these animals. In line with this suggestion, we found that only the high-frequency sensitivities (figure 3f) changed across metamorphosis when accounting for an increase in best frequencies (figure 3d), indicating that the two peaks observed in the vibrogram originate from two different end organs.

If assuming similar sensitivity of papilla hair cells in the juvenile and the adult axolotls, the reduction in vibration
sensitivity may originate from changes in the middle ear morphology across metamorphosis. Dissections and CT data from juvenile axolotls and adult tiger salamanders (figure 2) support the hypothesis that the columella is the functional element in the middle ear of aquatic juvenile salamanders, whereas the operculum after fusion of the columella to the otic capsule, and development of the opercularis muscle, is the functional element in the middle ear of the terrestrial adult tiger salamander [16,17]. The adult axolotls, however, apparently have both a movable columella and a functional opercularis system (figure 2; electronic supplementary material, figure S5). Comparing the vibration sensitivity with the morphology, our results suggest that the columella system and the opercularis system on their own may be equally efficient ways of coupling substrate vibrations to the amphibian papilla of the inner ear: juvenile axolotls and adult tiger salamanders had comparable sensitivity at high frequencies (figure 3d). The reduction in high-frequency vibration sensitivity found across metamorphosis in the axolotl (figure 3d,f), however, suggests that having both systems is less efficient. Thus, input through one system could be short-circuited by output through the other. Our results therefore seem to oppose the proposed functions of the opercularis system in aiding the transmission of substrate vibrations to the inner ear [17], at least in the axolotl. Collectively, the results suggest that the increased frequency range and vibration sensitivity at high frequencies of urodeles compared to those of animals with otolith auditory systems only, such as fish, are enabled by the additional structures of the urodele ear: the oval window containing movable inertial elements (columella and/or operculum) and the possession of the amphibian papilla (figure 1).

(c) Aerial hearing
Auditory systems of terrestrial animals are challenged by the large impedance mismatch between animal tissue and air, and therefore most of the sound energy is reflected when impinging on a terrestrial animal. Adult urodeles are atypical, and their auditory system therefore seems no more adapted to aerial hearing than fully aquatic vertebrates such as fish. Nevertheless, we confirm earlier indications of aerial hearing in adult urodeles [19] by showing that the adult axolotls and tiger salamanders are able to detect aerial sound with W-shaped audiograms and best sensitivity of approximately 80 dB re 20 μPa at 80 and 320 Hz (figure 3e). Surprisingly, no significant improvement in hearing sensitivities was found across the metamorphosis and so also the completely aquatic juvenile axolotls are able to detect aerial sound pressure with comparable sensitivity and frequency range to the adult salamanders (figure 3e). Again, evoked potential thresholds may be elevated 10–30 dB relative to actual thresholds and urodeles may therefore be able to detect sound pressures of approximately 50 dB re 20 μPa at high sound frequencies. This is comparable with the sound detection of atympanic frogs [47] (accounting for the difference in methodology) and atympanic reptiles [29], but is still relatively insensitive compared with the most sensitive tympanic anuran species [48]. The morphological and functional change in the urodele middle ear from the columella to the opercularis system [16,17] (figure 2) occurring during metamorphosis only had a minor effect on the aerial hearing of the axolotl (figure 3g). This is consistent with the fact that no morphological adaption to detection of aerial sound pressure develops in the middle ear across metamorphosis. The lack of increase in sensitivity found across metamorphosis is therefore likely to be rather a consequence of the relative good pressure sensitivity found in juveniles than of poor pressure sensitivity found in adult salamanders. In comparison, lungfish are also able to detect aerial sound [30], but only at low frequencies (less than 200 Hz) and at higher intensities than found here for salamanders. The possession of the oval window with movable inertial elements and the otoconia-free sensory epithelia in the inner ear therefore seem to enable urodeles to improve the sensitivity and frequency range when hearing in air despite being atypic. The lack of middle ear adaption for detection of aerial sound pressure, however, implies that urodeles are unable to detect sound pressure per se. Rather, urodeles may be hypothesized to detect sound-induced vibrations (figure 3d). When testing that notion, we found that sound-induced shaker vibrations were below vibration thresholds, and therefore unable to explain the sound pressure detection of the salamanders. Sound-induced head vibrations have previously been shown to be sufficient to explain sound detection in atympanic reptiles [29]. Use of the transfer function, from aerial sound pressure to head vibrations, determined for pythons with similar head size to the salamanders investigated here, suggests that urodeles detect aerial sound by detection of sound-induced head vibrations. Thus, the equivalent vibration thresholds calculated from salamander sound pressure thresholds (figure 3c) and python transfer functions [29] correspond to the vibration thresholds of the salamanders (figure 3d).

(d) Evolutionary perspectives
In many recent tetrapods, exemplified by most anurans, the impedance mismatch between air and animal tissue is overcome by the tympanic middle ear, which relays pressure-induced vibration of the tympanum via the middle ear ossicle to the endolymph of the inner ear (figure 1). There, fluid oscillations between the oval and the round window lead to hair cell deflection in papillae and otocia end organs by which the animal hears. By contrast, the auditory system of lungfish, the closest living relative of tetrapods [49], is regarded as primitively for tetrapods resembling the auditory system of the tetrapod ancestors [50]. The ears of lungfish can be characterized as unspecialized fish ears with a closed otic capsule and otocia end organs by which the animal hears. By contrast, the auditory system of lungfish, the closest living relative of tetrapods [49], is regarded as primitively for tetrapods resembling the auditory system of the tetrapod ancestors [50]. The ears of lungfish can be characterized as unspecialized fish ears with a closed otic capsule and otocia end organs by which the animal hears. By contrast, the auditory system of lungfish, the closest living relative of tetrapods [49], is regarded as primitively for tetrapods resembling the auditory system of the tetrapod ancestors [50]. The ears of lungfish can be characterized as unspecialized fish ears with a closed otic capsule and otocia end organs by which the animal hears.
tetrapods [53,54] suggest that the amphibian basilar papilla is homologous with that in the amniotes [55] and thus is a primitive character in tetrapods.

The morphology of the urodele auditory system resembles that of early ‘lepospondyl’ microsaur tetrapods [13,20], but it can be further regarded as an intermediate evolutionary stage between the primitive system of early tetrapods, as shown by lungfish and the tympanic ear of most amniors (figure 1). The auditory system of recent urodeles is therefore a relevant model for the auditory systems of early tetrapods before the evolution of the tympanic middle ear [13,20,52]. Caution should, of course, be taken when assuming that hearing of recent urodeles is representative of early tetrapods living some 300–350 Myr ago, but the morphological similarities between the urodele ear and the ears in early microsaur tetrapods lend support to the assumption of comparable auditory abilities.

We show that not only the terrestrial adult tiger salamanders, but also semi-terrestrial adult axolotls, and even completely aquatic juvenile axolotls, are able to detect airborne sound (figure 3c) despite their atypomanic middle ears. Our results hence suggest that the urodele auditory system, with an oval window containing free inertial elements, together with a papilla organ and the perilymphatic duct in the inner ear, enable them to have increased frequency range and sensitivity in air compared with fully aquatic vertebrates, such as fish. It follows from this suggestion that early tetrapods also may have been able to detect aerial sound before the appearance of the tympanic middle ear. This limited sensitivity may have provided the rudimentary hearing that, when selected for, led to gradual evolution of low-mass skin areas and bony structures that eventually formed the tympanic middle ear. Further, we show that urodeles are able to detect high-frequency sound pressure underwater (figure 3a).

This suggests that possession of air-filled structures enables pressure detection, as the lungs of urodeles are not mechanically connected to the inner ears. Hence, detection of underwater sound pressure may have appeared as a passive consequence of air breathing already in the aquatic ancestors of tetrapods. As the pressure-to-particle motion transduction of such gas-filled structures has the largest effect at frequencies above the resonance frequencies of the otolithic end organs, possession of such structures may have driven the evolution of high-frequency tuned hair cells. Accompanied by development of a lightweight, free inertial element in the oval window, this could have driven the evolution of free hair cell organs without an otolithic mass, a precursor of the basilar papilla, sensitive to high frequencies and responsible for aerial hearing in extant tympanic tetrapods. The evolutionary basis for pressure hearing could therefore have been formed already in water before the water-to-land transition. In concert, our results therefore imply a gradual change from particle motion detection in water to the pressure hearing on land, where high-frequency tuning in the aquatic air-breathing tetrapod ancestors and subsequent detection of sound-induced head vibrations in early tetrapods drove the evolution of aerial hearing, leading to the tympanic auditory systems of most modern tetrapods.

References


