Bilaterians usually possess a central nervous system, composed of neurons and supportive cells called glial cells. Whereas neuronal cells are highly comparable in all these animals, glial cells apparently differ, and in deuterostomes, radial glial cells are found. These particular secretory glial cells may represent the archetype of all (macro) glial cells and have not been reported from protostomes so far. This has caused controversial discussions of whether glial cells represent a homologous bilaterian characteristic or whether they (and thus, centralized nervous systems) evolved convergently in the two main clades of bilaterians. By using histology, transmission electron microscopy, immunolabelling and whole-mount in situ hybridization, we show here that protostomes also possess radial glia-like cells, which are very likely to be homologous to those of deuterostomes. Moreover, our antibody staining indicates that the secretory character of radial glial cells is maintained throughout their various evolutionary adaptations. This implies an early evolution of radial glial cells in the last common ancestor of Protostomia and Deuterostomia. Furthermore, it suggests that an intraepidermal nervous system—composed of sensory cells, neurons and radial glial cells—was probably the plesiomorphic condition in the bilaterian ancestor.

1. Background

The origin and evolution of animal nervous systems is controversially discussed [1–3]. In particular, it still remains an open question whether the complex centralized nervous system (CNS) found in the different clades of bilateral-symmetric animals (Bilateria) evolved independently out of a non-bilaterian nerve-net, or if this highly specialized bilaterian tissue shares a common origin [4–6]. The vast majority of species comprises two main clades within Bilateria: the deuterostomes, including all vertebrates, echinoderms, hemichordates, cephalochordates and tunicates; and the protostomes, including taxa such as annelids, molluscs, insects and nematodes. Most of these taxa have complex and CNS consisting of several neuron types with comparable molecular pathways, function and morphology [7,8]. Whereas most comparative neural investigations focus on different neuronal cell types supposed to be involved into various ways of stimulus perception and signal transmission, another important cell type also being part of bilaterian nervous systems is often neglected in comparative studies: the glial cells.

Glial cells are known to represent a class of non-neuronal supportive cells, constituting a common feature of the CNS in Bilateria. However, it still remains
controversial whether glial cells are homologous across deuterostome and protostome animals or evolved convergently in the bilaterian main clades [9]. Although supposed to be involved in almost all neural functions [10,11], these cells seem to be absent in several bilaterian taxa, as well as in non-bilaterian animals [9]. In particular, the epithelial nervous system of the Deuterostomia is occupied by a specific bipolar type of glial cells that span the entire thickness of the epidermis forming a regular ‘scaffold’, which are called radial glia [12]. In chordates, radial glial cells arise from neuroepithelial stem cells in the neural plate early during embryogenesis; they also represent neuronal stem cells that give rise to various types of other glial and neuronal cell types of the CNS [13–15]. Outside the Deuterostomia, however, the presence of radial glial cells has not yet been reported [16].

To examine the presence of radial glia-like cells also outside the Deuterostomia, we investigated taxa representing all major branches of the bilaterian tree and especially focused our study on non-deuterostomian groups possessing a supposedly plesiomorphic intraepidermal nervous system. By using an integrative approach including histology, transmission electron microscopy (TEM), immunolabelling and whole-mount in situ hybridization (WMISH), we show here that protostomes also possess radial glia-like cells, which are very likely to be homologous to those of deuterostomes. This implies an early evolution of radial glial cells in the last common ancestor of protostomes and deuterostomes (Nephrozoa). Moreover, our antibody staining suggests that the secretory character of radial glial cells is apparently maintained throughout their various evolutionary adaptations. So far, neuro- and gliogenesis originating from radial glial cells are only described for deuterostomes [13–15,17–19]. However, our results suggest that this cell type takes over important roles in nervous systems throughout Nephrozoa.

2. Material and methods

(a) Collection and fixation of specimens
For details, refer to the electronic supplementary material, table S1. Divergent collection and fixation details are specified where required.

(b) Larval rearing and culturing of Owenia fusiformis
Different larval stages of Owenia fusiformis were cultured under laboratory conditions and fixed as described previously [20].

(c) RNA-Seq and transcriptome assembly
Total RNA was extracted from cryofixed larvae using the Agen- court RNAAdvantage Tissue Kit (Beckman Coulter, Indianapolis, IN, USA). Library preparation and sequencing was performed by EMBL Genomics Core Facility (Heidelberg, Germany) using cation-based chemical fragmentation of RNA, Illumina Truseq RNA-Sample Preparation Kit and one lane of 100 bp paired-end read sequencing on Illumina HiSeq 2000. Raw reads were trimmed and error corrected with CUTADAPT 1.2.1 [21], the ErrorCorrectReads tool implemented in ALLPATHS-LG [22] and assembled with TRINITY [23].

(d) Gene cloning and RNA probe preparation
Contig sequences for the investigated genes were identified in the transcriptome dataset by bidirectional BLAST [24]. Whole transcripts or fragments were amplified by PCR with specific primers (Fw = AGTTTGGGATGCTTTGGG, Rv = TTTCCGCTACCGCTGG) from cDNA prepared with Superscript III (Invitrogen, Waltham, MA, USA), ligated into pgem-Teasy vector (Promega, Madison, WI, USA) and cloned into Top10 chemically competent Escherichia coli (Invitrogen). Clone sequences were verified by Sanger sequencing, DIG-labelled sense and antisense RNA probes were generated from plasmid DNA with T7- and SP6-RNA polymerases (Roche, Madison, WI, USA).

(e) Gene orthology
Reciprocal blast yielded unambiguous results for gene orthology assignment of Ofu-subcommisural organ (SCO). Furthermore, public databases (Genebank, JGI, Uniprot) and the transcriptome were screened for homologues by text search and BLAST with respective query sequences. For comparison with other members of the Thrombospondin-family, protein domains were analysed using SMART [25,26]. An overview is given in electronic supplementary material, figure S2. Accession number: MF358540.

(f) Immunohistochemistry, confocal laser scanning microscopy and image processing
Whole-mount preparations and vibratome sections were analysed. Specimens were anaesthetized in 7% MgCl2 in saltwater. For fixation, refer to electronic supplementary material, table S1. After rinses in 0.1 M phosphate-buffered saline (PBS) for at least 2 h, specimens were stored in PBS containing 0.05% NaN3 at 4°C. For vibratome sectioning, specimens were rinsed in 0.1 M PBS and embedded in gelatin/albumin medium. The blocks were cut with a VT1000S vibratome (Leica Microsystems, Wetzlar, Germany) into 80–100 μm thin sections. The sections were washed in PBS containing 0.1% Triton X-100 (PTA). Antibody staining was preceded by tissue permeabilization for 1 h in 0.1 M PBS containing 0.1% NaN3 and 0.1% Triton X-100 (PTA), suited by incubation in block-PTA (6% normal goat serum; Sigma-Aldrich, St Louis, MO, USA) overnight. For Hemichordata and Priapulida, PTA contained 2% Triton X-100. The primary antibody polyclonal rabbit antisemir against bovine RS (SCO-K10) (W. W Naumann, University of Leipzig, dilution 1:1000) was applied for 24–72 h in block-PTA. Afterw- 
s, specimens were rinsed in block-PTA and incubated with secondary fluorochrome-conjugated antibodies (goat anti-rabbit Alexa Fluor 488, Invitrogen, dilution 1:500) in block-PTA for 24–48 h. Subsequently, samples were washed in 0.1 M PBS (without NaN3), stained with DAPI for 15–30 min (5 mg ml−1 stock solution, final concentration 10 μg ml−1) and washed for 2 × 5 min in 0.1 M PBS. For experiments including larval and juvenile stages of O. fusiformis, specimens were rinsed for 2 × 5 min in PTW (PBS with 0.1% Tween 20) at room temperature (RT) and transferred into 10 μg proteinase K ml−1 PTW for 2–3.5 min depending on the developmental stage (24 hpf–3 dpf = 90 s; 7 dpf = 2 min; 14–21 dpf = 2.5 min; after metamorphosis = 3.5 min). After two short rinses in glycine (2 mg glycine ml−1 PTW), and 3 × 5 min washes in PTW, the specimens were refixed using 4% PFA in PBS containing 0.1% Tween 20 min at RT. Subsequently, the developmental stages were rinsed for 2 × 5 min in PTW and 2 × 5 min in THT (0.1 M Tris–Cl, 0.1% Tween), and blocked for 1–2 h in 5% sheep serum in THT according to the protocol of Conzelmann & Jékely [27]. The primary antibody polyclonal rabbit antisemir against bovine RS (SCO-K10) (W. W Naumann, University of Leipzig, dilution 1:1000) was applied for 24–72 h in THT containing 5% sheep serum at 4°C. Specimens were then rinsed for 2 × 10 min in 1 M NaCl in THT and 5 × 30 min in THT, and incubated subsequently with secondary fluorochrome-conjugated antibodies (goat anti-rabbit Alexa Fluor 488, Invitrogen proteins were stained with DAPI for 15–30 min–1
dilution 1:500) in THT containing 5% sheep serum for 24 h at 4°C. Subsequently, the samples were washed for 6 × 30 min in THT, stained with DAPI for 10–15 min (5 mg ml⁻¹ stock solution, working solution: 2 µl in 1 ml THT—final concentration 10 µg ml⁻¹) and washed for 2 × 5 min in THT. Samples were dehydrated in an ascending isopropanol series, treated in Murray’s clearing solution (benzyl alcohol plus benzyl benzoate, 1:2) and mounted between two coverslips in DPX (Sigma-Aldrich) or embeded in 90% glycerol/10% 10× PBS containing DABCO. Standard negative staining controls were performed for all antibodies, and in all cases, the omission of the primary and/ or secondary antibody resulted in no staining. Cross-reactivity analyses and immunoprecipitation for antigen characterization of the used SCO-K10 antibody were performed in earlier studies [28,29]. Samples were analysed with Leica TCS STED and Leica of the used SCO-K10 antibody were performed in earlier studies [28,29]. Samples were analysed with Leica TCS STED and Leica of the used SCO-K10 antibody were performed in earlier studies [28,29]. Samples were analysed with Leica TCS STED and Leica TCS SP5 microscopes (Leica Microsystems, Wetzlar, Germany). Z-stacks were processed with Leica AS AF v. 2.3.5 (Leica Microsystems) and IMARES 8.2 (Bitplane AG, Zurich, Switzerland). Final panels were designed using Adobe (San Jose, USA) PHOTOSHOP CC and ILLUSTRATOR CC.

(g) Whole-mount in situ hybridization
Larvae and juveniles of O. fusiformis were fixed for 2.5 h in 4% PFA in phosphate buffer with Tween (PTW; pH 7.4) and stored at −20°C in methanol until usage. The in situ hybridization procedure was performed as described previously [30] with some modifications: Proteinase K concentration was reduced to 5 ng ml⁻¹. Proteinase K treatment was 3.5 min for late larvae and 5 min for juveniles, hybridization buffer contained 5% dextran sulfate, the incubation time was 72 h and final staining was done with Fast Blue (Sigma-Aldrich, MO, USA). To evaluate staining, significance control experiments with sense probes were made. Furthermore, we combined in situ hybridization and antibody staining, by processing specimens after in situ hybridization with the immunohistochemistry procedure mentioned above.

(h) Ultrathin sections and transmission electron microscopy
Embedding was performed using the automated tissue processor (Leica Lynx Processor, Leica Microsystems GmbH, Saarland, Germany). After rinses of 4 × 20 min in 0.1 M PBS at 4°C, samples were treated for 2 × 60 min with 1% osmium tetroxide (Carl Roth, Karlsruhe, Germany) at 4°C. Afterwards, samples were rinsed for 20 min in 0.1 M PBS at 4°C and 20 min at RT. Subsequently, samples were dehydrated using an ascending acetone series at RT. The 70% acetone solution contained 1% phosphotungstic acid (Fluka, Sigma-Aldrich, Munich, Germany) and 1% uranyl acetate (Serva, Heidelberg, Germany) for post-contrasting. The samples were then gradually embedded at RT in non-hardening epoxy resin (Durecupan ACM Fluka; Sigma)-acetone mixtures (1:3, 1:1, 3:1; 60 min each), incubated in pure Durecupan and successively replaced by hardening Durecupan. After polymerization at 60°C for 2 days, semi-thin (500 nm) and ultrathin (60–65 nm) sections were cut with a microtome (Leica Ultracut UCT, Leica Microsystems, Saarland, Germany), employing a diamond knife. All sections were frontally orientated; displayed electron micrographs of O. fusiformis derived from the trunk of the worm, of Balanoglossus misakiensis from the posterior proboscis and from Asterias rubens from the isolated radial nerve.

For electron microscopy, ultrathin sections were transferred onto formvar-resin-laminated slot grids (Plano, Wetzlar, Germany) and post-contrasted with 3% uranyl acetate and 3% lead citrate. Sections were examined with a Sigma-0231 scanning electron microscope (27 kV; Zeiss), employing a STEM detector and ATLAS software (Zeiss).

(i) Histology and semi-thin sections
Balanoglossus misakiensis (Hemichordata) specimens were relaxed in 7% MgCl₂ in seawater for 5–10 min and fixed in ice-cold 2.5% glutaraldehyde in 0.05 M phosphate buffer + 0.3 M sodium chloride (pH 7.4). Primary fixation was stopped after 45 min with three rinses in buffer. Post-fixation with 2% OsO₄ was carried out for 30 min and stopped with three buffer rinses (15, 30, 30 min) followed by two rinses with ddH₂O (15, 30 min). After a graded series of ethanol specimens were embedded in Epon resin. Semi-thin sections (0.5 µm) of B. misakiensis (3 days post settlement) were carried out using a Leica Ultracut S. Sections were stained with toluidin blue and imaged with an Olympus BX-UCB mounted on an Olympus BX51 compound microscope.

Owenia fusiformis and Asterina gibbosa (collected in 2009 in Concarneau, France) were relaxed in a 7% MgCl₂ solution and fixed in Bouin’s fluid (modified after Dubosq-Basil) overnight. Adult specimens of Priapulus caudatus originated from a stock at the Natural History Museum of Denmark (ZMUC PRI-00119; collected in 1927 in Aggramiut, Greenland) and were fixed in Bouin’s fluid. All animals were dehydrated in an ascending ethanol series, incubated in methyl benzoate and butanol, preincubated in Histoplast (Thermo Scientific, Dreieich, Germany) at 60°C for 3 days with medium changes and embedded in Paraplast (McCormick Scientific, Richmond, VA, USA). Sections (5 µm) were made using a Reichert-Jung Autocut 2050 microtome (Leica, Wetzlar) and put on albumen–glycol-coated glass slides. According to a modified Azan staining method, sections were stained with Carmalaun, differentiated with sodium phosphotungstate (5%), washed in dH₂O, stained with aniline blue orange G and embedded with Malinol (Waldeck, Münster, Germany). In this staining, nervous system appears grey/reddish, nuclei of neuronal somata red and extracellular matrix blue. Musculature stains orange. Slices were analysed with an Olympus microscope (BX-51) and images taken with an Olympus cc12 camera equipped with the dot slide system (Olympus, Hamburg). Slices were aligned with imod and imod align (http://www.q-terra.de/biowelt/3drekon/guides/imod_first aid_pdf.pdf).

3. Results and discussion
We used histological staining, TEM investigations of ultrathin sections, immunolabelling and WISH to analyse the intra-epidermal nervous system of several taxa. Our antibody staining is based on the immunolabelling of SCO-spondin, using a polyclonal antibody (SCO-K10) derived from bovine Reissner’s substance and directed against bovine SCO-spondin [28,29]. This extracellular matrix glycoprotein is known to be expressed and secreted by the vertebrate SCO [31] and deuterostomian radial glial cells in general [32]. SCO-spondin is involved in early cell migration and axonal guidance, and seems to play important roles in vertebrate neurogenesis [33]. For our comparative investigations, we focused on taxa that represent the main branches of Nephrozoa [34] and exhibit an intraepidermal nervous system. In contrast with an intraepidermal plexus, an intraepidermal nervous system consists of one or several nerve cords that locally replace the epidermis [35]. This type of nervous system organization is presumably plesiomorphic for Bilateria [36]. Thus, we investigated the annelid O. fusiformis (Prostostomia, Lophotrochozoa), the priapulid P. caudatus (Prostostomia, Ecdysozoa) and the enteropneust
**Figure 1.** Phylogenetic hypothesis concerning Bilateria based on Cannon et al. [37], and SCO-K10-like immunoreactivity (-LIR) for selected taxa. Schematic images for radial glial cell-positive taxa are black, others are grey. Cross-sections of the nervous system of (a) Balanoglossus misakiensis, (b) A. rubens, (c) O. fusiformis and (d) whole mount of the anterior end of P. caudatus; z-projections; external epidermal = down and pointed by the white arrowhead (left and right in (d)). SCO-K10-LIR reveals the presence of elongated cells (yellow arrowhead) within epidermis. The yellow dotted line marks the position of the nerve cord in (c). Scale bars, 20 μm (a–c); 5 μm (d). Schematic images from phylopic.org.

B. misakiensis (Deuterostomia, Hemichordata) (figure 1). For comparisons, we re-examined the sea star A. rubens (Deuterostomia, Echinodermata) (figure 1), where the here-applied antibody had already been tested successfully [29].

The oweniid O. fusiformis is part of the earliest-branching split of annelids [39]. We found that the ventral nerve cord of this species is situated intraepidermally (figure 2a,b), thereby confirming results of an anatomical study on another oweniid species [40]. A higher magnification reveals the presence of distinct, regularly arranged structures traversing the entire epidermis from the basal lamina towards the epithelial surface. Examination of the same region using an antibody directed against SCO-spondin reveals prominent SCO-K10-like immunoreactivity (-LIR) in these cells (figure 1a). In fact, the ventral cord and brain of O. fusiformis exhibit radially arranged SCO-K10-LIR and distinct gene expression of Ofu-SCO-spondin, crossing epidermis and intraepidermal nervous system (figures 1c and 3a–f–j–n). Notably, a spatial separation of the SCO-protein in the apical cell area and of the Ofu-SCO mRNA in basal parts of the cell is observable (figure 3g–i). Alignment of immunohistochemical, histological and ultrastructural investigations (figure 3j–n) confirms the exclusive presence of radially arranged cells traversing the intraepidermal nerve cord and the adjacent epidermis, featuring SCO-K10-LIR, exhibiting prominent intermediate filaments (figures 4b and 5c) and flanking neuronal cells with synaptic vesicles (figure 5c). These cells possess funnel-shaped endfeet terminating at the basal lamina (figure 4a) and microvilli penetrating the cuticle at the opposite surface (figure 4c,e). The apical end of the cells contains secretory vesicles (figure 4d) and cilia (figure 5d,e). Somata of glial and neuronal cells are located in the apical zone of the epidermis (figure 5c).

A comparable situation is found in the enteropneust B. misakiensis. This species also exhibits an intraepidermal nervous system (figure 2e,f). Antibody staining of SCO-spondin reveals numerous radially arranged structures in the posterior end of the proboscis in early juveniles. These structures span the entire epidermis in the baso-apical direction (figure 1a). Notably, the SCO-K10-LIR is restricted to the posterioridorsal part of the proboscis, where the intraepidermal proboscis plexus is located. Observations using TEM show that the somata of the cells mostly reside in the apical section of the epidermis (figure 5g,h). The cells also contain prominent intermediate filaments and funnel-shaped endfeet abutting the basal lamina at the inner surface, as well as distinct apical microvilli and cilia at the apical cell surface (figures 4f,g and 5h,i). This result reveals, for the first time, the existence of radial glial cells—and glial cells in general—in Hemichordata [9]. The intraepidermal nervous system of the echinoderm A. gibbosa is also characterized by prominent radially arranged cells traversing the entire intraepidermal nerve cord and the adjacent epidermis in adult specimens (figure 2c,d). In agreement with a previous study [29], immunohistochemical analysis unveils distinct SCO-K10-LIR displayed by these cells in A. rubens (figure 1b). Ultrastructural investigation of the nerve cord in both asteroidean echinoderm species shows the presence of dense bundles of intermediate filaments within these cells (figures 4i and 5k,l). Further on, these cells traverse the...
entire epithelium with their somata mostly located in the apical neuroepithelium (figure 5k,l), and bear prominent funnel-shaped basal endfeet at the basal lamina (figure 4i), as well as apical microvilli and cilia at the apical cell surface (figures 4j and 5m,n). This is in accordance with earlier findings [41]. Using the same antibody, evidence for the presence of radial glial cells has also been reported for the nervous system of holothurian echinoderms [32,42].

Literature screening furthermore revealed the presence of cells (called ‘tanycytes’) with radial glia-like alignment and ultrastructure in the ecdysozoan Scalidophora (Priapulida, Loricifera, Kinorhyncha; see also [43] and figs 1 and 3b in [44] for further ultrastructural details). This prompted us to study the intraepidermal nervous system of P. caudatus (Priapulida) (figure 2g,h). Indeed, we found radially aligned SCO-K10-LIR in cells that span the thickness of the intraepidermal nerve cord and the adjacent epidermis in larvae of P. caudatus (figure 1d).

The ultrastructures of the intraepidermal nervous system of all studied protostomes and echinoderms resemble each other in detail. As the central nervous system of vertebrates ontogenetically originates from epithelia [45], the resemblance of the intraepidermal nervous systems of echinoderms, hemichordates and protostomes provides further support for the homology of epithelial nervous systems in Bilateria, which probably represents the plesiomorphic condition. Within this intraepidermal nervous system, radial glia represent an important non-excitable structural and functional component, and are therefore proposed to be a key feature of all epithelial nervous systems.

Combining all evidence, including the shared topological position and similar ultrastructural composition, the

Figure 2. Semi-thin sections of the neuroepithelium in (a,b) O. fusiformis, (c,d) A. gibbosa, (e,f) B. misakiensis and (g,h) P. caudatus. Azan (a–d,g,h) and toluidine-blue (e,f) staining. Azan stains somata and cell processes reddish, neuropil grey, nuclei red and extracellular matrix blue. Musculature is orange. Toluidine-blue stains somata, cell processes and extracellular matrix in different blue intensities. (a,e) The intraepidermal nervous system (is) locally replaces the epidermis (ep) (bl, basal lamina; in, intestine; mo, mouth; pb, proboscis). (b,d,f,h) Higher magnification of regions indicated by dotted line in (a), (c), (e) and (g), respectively (no scale bar included). Arrows indicate radial structures in (b) and (d). Scale bars, 100 μm.
SCO-spondin-LIR, the expression of the Ofu-SCO mRNA and the early developmental occurrence, all data strongly support the conclusion that the cells we located in oweniid annelids, as well as in enteropneusts, echinoderms and priapulids, are homologous to the radial glial cells described in chordates. This hypothesis of homology is based on positional and structural homology criteria [46]. Using a comparative approach [36], this implies the evolution of this cell type in the last common ancestor of Nephrozoa (Protostomia and Deuterostomia).

Noteworthily, radial glial cells are absent in the subepidermal ganglia (sensu Richter et al. [35]) of the protostomian CNS. Although this difference might suggest a convergent
evolution of glial cell types, from different origins, in the Protostomia and Deuterostomia, one should keep in mind that radial glia are missing in the ganglia of the peripheral nervous system of vertebrates as well. This is simply because their existence requires both a basal and an apical cell surface, which are present only in an epithelium [12].

Several studies across all major chordate and echinoderm taxa showed that radial glia are secretory [32,38,42,47,48]. In our study, we provided evidence for a secretory protein (SCO-spondin) expressed in all species studied. So far, secretion of SCO-spondin, which is the main component of the so-called Reissner’s substance, is best characterized in the vertebrate cerebrospinal fluid [49], which is present in the central canal of the chordate dorsal nerve cord [50]. Notably, Reissner’s fibre is secreted only by a specialized type of glial cells, the radial glia [29,47]. The function of Reissner’s fibre has been suggested to participate in the regulation, circulation and production of cerebral spinal fluid of vertebrates [51–53]. Reissner’s fibre is also known from tunicates and cephalochordates [54], and it has recently been shown that vertebrate SCO-spondins have indeed orthologues in invertebrate taxa [55]. However, no Reissner’s fibre-like structure has been reported outside Chordata. Interestingly, Arendt et al. [56] recently hypothesized two major morphological innovations that contributed to set the stage for the evolution of the animal nervous systems: (i) the establishment of a mucociliary sole enabling extracellular digestion in gastric cavities followed by the evolution of a nerve-net, and (ii) folding of the inner surface of these animals.

Figure 4. Ultrastructure of glial components in the neuroepithelium of (a–e) O. fusiformis, (f–h) B. misakiensis and (i–k) A. rubens. (a,f,i) Funnel-shaped basal processes (including endfeet) of radial glial cells (arrowheads) terminate at inner epithelial surface (bl, basal lamina). (b,g,j) Inner cell processes of radial glial cells (arrowheads) traverse whole epithelium up to apical nuclear layer; asterisks indicate the location of glial cell somata. (c,e,h,k) Microvilli (mv) from radial glia protrude and break through cuticle (cu). (d) Presumable radial glial cell during release of secretory products into the cuticle. Scale bars, 1 μm.
into metameric series of gastric pouches, which optimized nutrient provision, thereby allowing larger body plans, subsequent evolution of bilateral symmetry and the evolution of specialized nervous subsystems. Strikingly, in this context, Reissner’s fibre is interpreted as a possible remnant of a mucociliary sole. Our investigations support a secretion of the Reissner’s fibre component SCO-spondin in several non-chordate taxa. Thus, the secretory character of radial glial cells appears to be maintained throughout their various evolutionary adaptations, providing a mucociliary sole, components of the cuticle or Reissner’s fibre.

In vertebrates, it is well known that radial glial cells have a major impact on early neuronal development. So far, neuro- and gliogenesis originating from radial glial cells are only described for chordates and, partially, for echinoderms [13–15,17–19,57]. However, as our comparative investigation suggests the presence of radial glial cells in the last common ancestor of Nephrozoa, it is tempting to speculate about a possible homology of a neuronal stem cell system across nephrozoan taxa in general. Future studies focusing on neuro- and gliogenesis in protostomes with secretory radial glial cells will help to elucidate this hypothesis.

Data accessibility. The figures and table supporting this article have been uploaded as the electronic supplementary material.

Authors’ contributions. C.H., A.K., A.R. and C.B. conceived the study, analysed the data and drafted the manuscript. C.H., A.K., P.B., S.K.-S., E.U., I.K., H.K., H.H. and T.B. performed experiments, contributed specimens and/or raw data. All authors read and approved the final version of the manuscript.

Competing interests. We declare we have no competing interests.

Funding. C.H. was financed by a personal research fellowship from the DFG (HE 7224/1-1). A.K. was financially supported by the
Acknowledgements: We thank José M. Martin-Durán and Andreas Heinzel (University of Bergen, Norway) for providing priapulid Iorcia larvae, and Martin V. Sørensen (Natural History Museum, Denmark) for adult priapulids. We kindly acknowledge W. W. Naumann who provided the SCO-K10 antibody and Judith Craatz (University of Leipzig, Germany) for technical assistance.

References


