Identification of an ant queen pheromone regulating worker sterility

Luke Holman1*, Charlotte G. Jørgensen2,3, John Nielsen2 and Patrizia d’Ettorre1,†

1Centre for Social Evolution, Department of Biology, University of Copenhagen, Universitetsparken 15, 2100 Copenhagen, Denmark
2Department of Life Sciences, Bioorganic Chemistry, University of Copenhagen, Thorvaldsensvej 40, 1871 Frederiksberg, Denmark
3Department of Medicinal Chemistry, University of Copenhagen, Jagtvej 162, 2100 Copenhagen, Denmark

The selective forces that shape and maintain eusocial societies are an enduring puzzle in evolutionary biology. Ordinarily sterile workers can usually reproduce given the right conditions, so the factors regulating reproductive division of labour may provide insight into why eusociality has persisted over evolutionary time. Queen-produced pheromones that affect worker reproduction have been implicated in diverse taxa, including ants, termites, wasps and possibly mole rats, but to date have only been definitively identified in the honeybee. Using the black garden ant Lasius niger, we isolate the first sterility-regulating ant queen pheromone. The pheromone is a cuticular hydrocarbon that comprises the majority of the chemical profile of queens and their eggs, and also affects worker behaviour, by reducing aggression towards objects bearing the pheromone. We further show that the pheromone elicits a strong response in worker antennae and that its production by queens is selectively reduced following an immune challenge. These results suggest that the pheromone has a central role in colony organization and support the hypothesis that worker sterility represents altruistic self-restraint in response to an honest quality signal.

Keywords: social insect; cuticular hydrocarbon; queen signal; Lasius niger; handicap

1. INTRODUCTION

Worker sterility is the defining feature of eusociality, and is therefore fundamental to any explanation of its evolutionary origin and maintenance. The degree to which worker sterility is driven by cooperation or conflict (Lehmann & Keller 2006; Ratnieks et al. 2006; Boomsma 2009) and individual- or colony-level selection (Keller 1999; Wilson & Hölldobler 2005; Okasha 2006) remain active areas of research. At the proximate level, the genetic (Grozinger et al. 2003; Nelson et al. 2007; Schwander & Keller 2008; Alaux et al. 2009; Wurm et al. 2010) and developmental (Khila & Abouheif 2008; Roat & Landim 2008; Johnson & Linksvayer 2010) bases of reproductive division of labour have been elucidated with increasing resolution, although the systems that determine when and why individuals relinquish sterility and switch to individual reproduction are less well understood.

Queen-produced pheromones that maintain worker sterility are thought to be taxonomically widespread, as queens, their eggs and queen-derived chemicals have been shown to reduce or eliminate worker reproduction, and because queens typically produce chemicals that are absent or minimally expressed in workers (e.g. Vargo 1992; Peeters et al. 1999; Dietemann et al. 2003; Cuvillier-Hot et al. 2004a; Endler et al. 2004; Monnin 2006; Dengler-Crish & Catania 2007; Korb et al. 2009; Bhadra et al. 2010). However, the honeybee is the only insect in which primer pheromones (i.e. pheromones with a physiological effect) have been definitively identified (Le Conte & Hefetz 2008), meaning that it is difficult to draw general conclusions about the factors regulating sterility. Queen pheromones underpin the proximate and ultimate causes of worker sterility: in the honeybee, they cause changes in worker gene expression (Grozinger et al. 2003; Beggs et al. 2007) and physiology (Kaatz et al. 1992; Beggs et al. 2007) that mediate the transition from indirect to individual reproduction, and they have been postulated to be either a manipulation that is detrimental to workers (‘queen control’) or a signal to which workers are selected to respond (‘queen signal’; Keller & Nonacs 1993; Heinze & d’Ettorre 2009). Queen pheromones are also interesting because they are thought to be central to the colony’s ‘social physiology’, the superorganismic analogue of regulatory mechanisms such as hormones (Johnson & Linksvayer 2010). Elucidating the identity, modus operandi and fitness consequences of queen pheromones in additional taxa is therefore likely to produce new insights into social evolution.

Here, we identify a multi-functional queen pheromone from the black garden ant Lasius niger. In a previous study, we found that the cuticular hydrocarbon 3-methylhentriacontane (3-MeC31) was strongly correlated with queen productivity, maturity and likelihood
of avoiding execution by workers in colonies with supernumerary queens (Holman et al. 2010). The results of Holman et al. (2010) imply that 3-MeC₃₁ is a signal of queen quality, and verbal models have suggested that the queen pheromones hypothesized to regulate reproductive division of labour are likely to be honest quality signals (Keller & Nonacs 1993; Zahavi & Zahavi 1997; Heinze & d’Ettorre 2009; van Zweden 2010). We therefore developed a novel synthetic pathway for 3-MeC₃₁ in order to test whether it (i) affects worker ovarian activation, (ii) influences worker behaviour and (iii) is detectable by workers. We also quantified queens’ chemical profiles after an experimental immune challenge, to assess whether 3-MeC₃₁ could provide information on queen condition to workers. Lastly, we found that 3-MeC₃₁ is abundant on the surface of queen-laid eggs, giving insight into its function and mode of action.

2. MATERIAL AND METHODS

(a) Comparison of worker, queen and egg chemical profiles
Cuticular hydrocarbons were extracted from L. niger queens (n = 20) and analysed as previously described (Holman et al. 2010). In short, cuticular hydrocarbons were extracted for 10 min in 150 µl pentane; the pentane was allowed to evaporate, and the extract was re-diluted in 60 µl pentane. We then injected 2 µl of extract into the GC-MS using an auto-sampler. Analysis of egg and worker surface hydrocarbons was the same except for the extraction and injection methods; 10 eggs or one worker were placed in a 200 µl glass insert and extracted for 3 or 10 min, respectively, in 20 µl pentane, 2 µl of which was then manually injected into the GC-MS (n = 20). Peak areas were analysed using multivariate statistics (using transformed data as in Holman et al. 2010) and univariate statistics (using proportion data and GLMs).

(b) Synthetic cuticular hydrocarbons
Synthesis of 3-MeC₂₇ and 3-MeC₃₁ is described in the electronic supplementary material. C₉ and C₃₁ were purchased from Sigma-Aldrich.

(c) Effects of 3-MeC₃₁ on worker ovarian activation and behaviour
Lasius niger workers were collected from six wild colonies in Copenhagen, Denmark. Collected workers were divided into three equal groups, each of which was given a model queen made from the tip of a glass vial. Every 12 h for 37 days after collection, the model queen was removed, coated with 10 µl of a pentane solution of (i) 0.01 µg µl⁻¹ 3-MeC₃₁, (ii) 0.01 µg µl⁻¹ hentriacontane (C₃₁) or (iii) pentane only, and replaced once completely dry (blind, using a labelling code). The alkane C₃₁ was chosen as a control hydrocarbon because it is also a queen-type cuticular hydrocarbon of L. niger (electronic supplementary material, table S1) and has the same chain length, but was previously found to be unrelated to queen productivity, maturity or survival (Holman et al. 2010). Highly purified HPLC-grade pentane (Sigma-Aldrich) was used throughout.

After 37 days, all colonies were frozen for dissection. Ovarian activation was scored on a scale of 1–4: (1) completely empty; (2) one or two very small eggs and/or developing nurse cell material; (3) one to three developing eggs in both ovarioles or large eggs in one ovariole; and (4) well-developed eggs in both ovarioles (blind, using a different labelling code to the behavioural observations). Production of males by workers occurs in natural colonies of L. niger (Fjerdingstad et al. 2002), although oviposition was not observed in our small laboratory colonies.

On days 3–37 of this experiment, we conducted 3 min of behavioural observations (blind to treatment) starting 10 s after the replacement of the model queen, with the aid of ETHOLOG v. 2.2.5 software (Ottoni 2000). We recorded aggression towards the model queen (duration of attack multiplied by number of workers attacking) and the number of aggressive worker–worker interactions. Ovary and behavioural data were analysed with quasi-Poisson GLMMs with colony (and observation day, for the behavioural observations) as a random factor, in order to account for non-normal errors, overdispersion and within-colony similarity.

(d) Electroantennography of synthetic hydrocarbons
We collected workers from a wild colony and used them within 6 h in electroantennography (EAG) trials (protocol adapted from d’Ettorre et al. 2004). The left antennal flagellum was excised (n = 25 workers) and mounted between two pulled glass capillaries containing insect Ringer, which bathed two Ag–AgCl electrodes. The electrode holding the proximal end of the flagellum was connected to a ground wire, while the other was connected via an amplifier to a signal acquisition interface board (IDAC; Syntech, Hilversum, The Netherlands) for signal transfer to a PC. The antenna was placed in a stream of purified, humidified air, and the amplitude of the depolarization response of the antennal neurons was recorded in millivolts (using EAG 2000 software; Syntech) following exposure to six different stimuli: a pentane control, and pentane solutions (all 0.5 µg µl⁻¹) of C₂₀, C₃₁, 3-MeC₂₇ and 3-MeC₃₁, as well as a mixture containing all of these hydrocarbons. These hydrocarbons are all present on the cuticle of queen L. niger (electronic supplementary material, table S1), but all had a non-significant or weak (relative to 3-MeC₃₁) relationship with queen fertility, maturity and survival (Holman et al. 2010).

We placed 10 µl of hydrocarbon solution on a 5 × 15 mm piece of filter paper in a new Pasteur pipette heated to 70 °C on a hotplate, and immediately blew a pulse of air through the pipette onto the flagellum. Before starting each run, we blew a single pulse of air onto the antenna to verify that it was responsive. The treatment order was randomized, and the experiment was conducted and analysed blind. Responses were standardized against the control for each antenna by setting the response to the pentane control as 100 and transforming the other treatments accordingly. The data were analysed using a GLMM with Gaussian errors and antenna as a random factor, in order to account for non-normal errors, overdispersion and within-colony similarity. (d) Effects of immune challenge on production of 3-MeC₃₁ by queens
Lasius niger queens were collected during a mating flight in Copenhagen and allowed to mature and rear workers in the laboratory for 201 days. To administer an immune challenge, we starved queens for 24 h (Moret & Schmid-Hempel 2000)
and then pierced their inter-pleural membranes using a sterilized pin coated with either 2.5 mg ml\(^{-2}\) lipopolysaccharide (Sigma-Aldrich) in sterile Ringer, or Ringer alone (blind and randomized). Queens were isolated from their colonies for 24 h then frozen for cuticular hydrocarbon analysis as described above; the peak areas were analysed blind. All statistical tests were performed in R v. 2.8.1 and validated using diagnostic plots.

3. RESULTS

(a) 3-MeC\(_{31}\) is a major component of queen and egg chemical profiles

The cuticular hydrocarbon profiles of queens and workers were markedly different (discriminant analysis based on six principal components explaining 85% of the variance: Wilk’s \(\lambda = 0.03, F_{6,33} = 155, p < 0.0001\)), with 3-MeC\(_{31}\) showing the strongest caste specificity (queens had a 6.1 times higher proportion than workers; figure 1a; electronic supplementary material, table S1) and being the most abundant single compound in the queen profile. 3-MeC\(_{31}\) was also the most abundant hydrocarbon on the surface of queen-laid eggs (figure 1a; electronic supplementary material, table S1).

(b) Synthetic 3-MeC\(_{31}\) reduces worker ovarian activation and aggressive behaviour

After 37 days of separation from the queen, worker ovarian activation was significantly lower in colony fragments that had been supplemented twice daily with synthetic 3-MeC\(_{31}\) rather than pentane solvent (figure 1b; GLMM: \(t = 2.76, p = 0.006, n = 478\) workers) or the control hydrocarbon C\(_{31}\) (\(t = 2.27, p = 0.024\)). Ovarian activation did not differ between C\(_{31}\) and pentane-treated workers (\(t = 0.64, p = 0.52\)). This experiment was replicated with workers from six colonies; there was no significant treatment\(\times\)colony interaction term (\(F\)-test comparing models with colony fitted as a fixed factor: \(F_{9,260} = 0.95, p = 0.47\)), showing that the effect of 3-MeC\(_{31}\) was consistent across colonies (electronic supplementary material, figure S1). We therefore conclude that 3-MeC\(_{31}\) is a primer pheromone that negatively affects the activation of worker ovaries.

Workers frequently attacked the glass model queens to which we applied the hydrocarbon solutions. However, models coated with 3-MeC\(_{31}\) were attacked significantly less than those treated with C\(_{31}\) (GLMM: \(t = 5.83, p = 0.03\)). The duration of attack did not differ between the pentane and C\(_{31}\)
random factor: p significantly higher (starred; workers in the form of body-jerking threat displays, observations). We also recorded mild aggression among treatments (t = 1.16, p = 0.25) (figure 1c; n = 630 observations). We also recorded mild aggression among workers in the form of body-jerking threat displays, although the number of aggressive acts did not differ between treatment groups (p > 0.1; n = 630). There was therefore no evidence that the onset of worker ovarian activation was accompanied by increased worker–worker aggression as in some other social insects (e.g. Cuvillier-Hot et al. 2004a; Korb et al. 2009).

(c) 3-MeC31 elicits a strong electrophysiological response in worker antennae

In EAG trials, 3-MeC31 induced a stronger response in excised worker antennae than the pentane control (figure 2; mixed-effects model with antenna as a random factor: t = 2.04, n = 25, p = 0.043), demonstrating that the queen pheromone is detected by worker antennae. None of the other synthetic hydrocarbons tested (C29, C31 and 3-MeC27) induced a significantly greater antennal response than the control (all p > 0.46). The response to 3-MeC31 was significantly higher than that to C29 (t = 2.33, p = 0.048) and C31 (t = 2.75, p = 0.020), and non-significantly higher than that to 3-MeC27 (t = 1.31, p = 0.22) and the mixture of all four hydrocarbons (t = 0.92, p = 0.35).

(d) Immune challenge reduces the amount of 3-MeC31 present on queen cuticle

The cuticular hydrocarbon profiles of queens subjected to a lipopolysaccharide immune challenge were significantly different from those of controls (discriminant analysis based on eight principal components explaining 86% of the variation; Wilk’s λ = 0.60, χ² = 16.9, p = 0.031), primarily because the mean proportion of the chemical profile composed of 3-MeC31 was 21% per cent lower in challenged queens (figure 1f; ANOVA: t₉₈ = 2.83, p = 0.007). The proportions of all 29 of the other hydrocarbon peaks did not significantly change following immune challenge (all p₉₈ > 0.06).

4. DISCUSSION

These experiments demonstrate that 3-MeC31: (i) is a primer pheromone that negatively affects worker ovarian activation; (ii) is also a releaser pheromone (i.e. one that affects behaviour) that is perceptible to workers and influences aggressive behaviour; (iii) elicits a comparatively strong response in worker antennae, implying the presence of many olfactory receptor neurons sensitive to 3-MeC31, consistent with its function as a pheromone; and (iv) displays condition-dependent expression. We also found that 3-MeC31 is the major component of the chemical profile of queen-laid eggs.

To our knowledge, 3-MeC31 is the first insect primer pheromone to be definitively identified outside of the honeybee (Le Conte & Hefetz 2008), and the first cuticular hydrocarbon demonstrated to affect conspecific reproductive physiology in any species. Primer pheromone activity of queen cuticular hydrocarbons is nevertheless likely to be common throughout the social insects. Differences in the cuticular hydrocarbon profiles of queens and workers have been reported in many species of ants, bees, wasps and termites (e.g. Peeters et al. 1999; Dietemann et al. 2003; Monnin 2006; Sramkova et al. 2008; Liebig et al. 2009; Peeters & Liebig 2009). Moreover, in Camponotus floridanus ants, workers in queenless colonies do not reproduce while queen-laid eggs are present (Endler et al. 2004); the eggs are coated with a hydrocarbon mixture similar to the cuticle of queens, consistent with regulation of worker sterility by one or more hydrocarbons. Similarly, queen corpses reduce the reproductive output of live queens in Solenopsis invicta ants (Vargo 1992), and in the wasps Polistes gallicus and Ropalidia marginata queens apparently prevent subordinate reproduction with chemicals from an abdominal gland (Dappporto et al. 2007) and Dufour’s gland (Bhadra et al. 2010), respectively. Interestingly, R. marginata queens had more 3-MeC31 in Dufour’s gland than did workers, suggesting that this compound may act as a primer pheromone in distantly related species. Cuticular hydrocarbons have also been shown to regulate reproduction indirectly through their role in ‘worker policing’: illegitimate reproductives are identified by their cuticular hydrocarbons and aggressed by their nest-mates (e.g. Peeters & Liebig 2009; Smith et al. 2009).

In honeybees, queen pheromones are detected via an odorant binding protein in workers’ antennae (Wanner et al. 2007), leading to reductions in juvenile hormone titre (Kaatz et al. 1992), dopamine production (Beggs et al. 2007) and dopamine receptor gene expression (Beggs et al. 2007) that cause workers to remain sterile. Available data suggest that the physiological mechanisms by which 3-MeC31 affects worker reproduction in L. niger are probably very similar. Firstly, our results demonstrate that workers perceive 3-MeC31 via their antennae. A recent study of L. niger found a gene (Ln385.5) with worker-biased expression that encodes a homologue of the pheromone-binding protein ASPI (Graff et al. 2007). This protein is found in the antennal olfactory sensillae of worker and drone honeybees, where it binds to queen pheromone (Danty et al. 1999), so it is possible that Ln385.5 is also involved in the perception of queen pheromones. Secondly, juvenile hormone also regulates ovarian activation in L. niger.
(Sommer & Hölldobler 1995), and in S. invicta loss of the dominant queen affects the expression of genes that regulate juvenile hormone levels in subordinate queens, causing them to activate their ovaries (Wurm et al. 2010).

Our immune challenge experiment indicates that 3-MeC31 provides information about a queen’s immune status or overall condition. This result implies that production of 3-MeC31 is physiologically costly relative to other cuticular hydrocarbons (assuming that pheromone production is always beneficial to queens), and therefore supports the prediction that queen pheromones should only be evolutionary stable when they honestly signal a queen’s reproductive potential (Keller & Nonacs 1993), because costly traits act as handicaps that constrain dishonest signalling (Johnstone & Grafen 1993; Zahavi & Zahavi 1997; Heinze & d’Ettorre 2009; van Zweden 2010). Several other lines of evidence suggest that putative social insect queen pheromones are handicaps. Queen-specific cuticular hydrocarbons are typically methylated alkanes or alkenes, which are thought to confer inferior protection against desiccation compared with the hypothetically ancestral compounds, linear alkanes (Monnin 2006; Le Conte & Hefetz 2008). Also, reproductive development in insects is correlated with hormone titres (Heinze & Schrempf 2008) as well as surface chemicals; hormones influence condition and survival, for example through effects on immune function (Rolf & Siva-Jothy 2002) and anti-oxidant activity (Heinze & Schrempf 2008). Therefore, the costs of pheromone synthesis might arise from the physiologically expensive hormone levels required for their production (as suggested for sexual signals; Folstad & Karter 1992), although further biochemical data are required to test this hypothesis. First steps in this direction have been achieved by studies supplementing queens with juvenile hormone analogues, which suppress reproduction; hormone treatment was associated with a reduction in reproductive-like chemicals in queenless ants (Cuviillier-Hot et al. 2004b), but not in honeybees (Malka et al. 2009). Cuticular hydrocarbons associated with reproductive activity can also attract aggression in certain contexts (e.g. when expressed by individuals with relatively low reproductive potential), and thereby incur costs (Peeters & Liebig 2009; Smith et al. 2009).

An alternative to the handicap hypothesis of honest queen pheromones is based on the ‘index’ concept (sensu Maynard Smith & Harper 2003); pheromone production might be inextricably linked to reproductive physiology (e.g. by shared dependence on common biosynthetic pathways), so that dishonest signalling is impossible (Heinze & d’Ettorre 2009; Smith et al. 2009; van Zweden 2010). Our immune challenge data are also consistent with this hypothesis; immune activation might have depressed queens’ reproduction, which in turn lowered pheromone production. Future studies will need to investigate the costs, genetics and biochemistry of pheromone production in order to distinguish between these hypotheses.

As well as being abundant on the cuticle of queen L. niger, 3-MeC31 is the most plentiful hydrocarbon on the surface of queen-laid eggs, and is also found on cocoons (Holman et al. 2010). Being present on brood may increase the frequency with which workers encounter 3-MeC31, which has very low volatility; ant brood has been shown to inhibit sexual production (Edwards 1987) and worker oviposition (Endler et al. 2004), suggesting that eggs are a means of distributing queen pheromones. Many L. niger cuticular hydrocarbons, including 3-MeC31, are also present on the nest soil (Lenoir et al. 2009), which could be another mechanism of dispensing the signal. Another function of brood-borne 3-MeC31 may be the regulation of queen productivity via negative feedback. Lasius niger queen productivity was lower in the presence of brood and other queens (Holman et al. 2010), so it is possible that 3-MeC31 affects the reproductive state of queens as well as workers (although the dose–response curve of the two castes would probably be different). The presence of the queen pheromone on eggs may also contribute to ensuring that the pheromone is an honest signal of fertility, because fertile queens will produce more eggs and thereby expose workers to greater quantities of pheromone. Lastly, 3-MeC31 might serve as an egg-marking signal used by workers to decide which eggs to rear (e.g. Endler et al. 2006; van Zweden et al. 2009).

Together with previous work, our results show how condition-dependent queen pheromones could act as parsimonious ‘master signals’ at the centre of the colony’s social physiology that quantitatively modulate multiple colony-level traits. If pheromone production declines with the condition of the queen (e.g. when the queen becomes old or ill), worker behaviour associated with the absence of the queen may be initiated before the queen dies. Where the same pheromones are present on brood (Endler et al. 2004; van Zweden et al. 2009), declining brood number may similarly contribute to these worker responses (Edwards 1987; Endler et al. 2004), as well as allowing queens to tune their reproductive rate to the current number of brood as mentioned above. If the queen pheromone also affects worker aggression (as implicated here and in several other ants; Vander Meer & Alonso 2002; Peeters & Liebig 2009; Smith et al. 2009; Moore & Liebig 2010; Wurm et al. 2010), the pheromone could also be used by the colony to decide who should reproduce. In L. niger, colonies are often co-founded by multiple queens, but only one queen survives after the first workers eclose (Sommer & Hölldobler 1995). A queen’s likelihood of being spared execution by workers is correlated with the amount of 3-MeC31 on her cuticle, implying that workers use this chemical to selectively kill the least fertile queens (Holman et al. 2010). Queen-like hydrocarbons also facilitate identification and punishment of reproductive workers in the ant Aphaenogaster cockerelli (Smith et al. 2009) and are thought to signal reproductive rank in queenless ants with dominance hierarchies (Peeters & Liebig 2009). A queen-derived cue also modulates worker aggression and consequently adoption of new queens in S. invicta (Vander Meer & Alonso 2002), and chemicals from the queen’s sting gland prevent subordinate queens from shedding their wings and becoming reproductive (Vargo 1997). In the honeybee, production of most queen pheromone components is lower in ‘drone-producing’ and virgin queens relative to fully fertile queens (Strauss et al. 2008), resulting in a reduced behavioural response by workers (Kocher et al. 2009), and pheromone quantity or quality deteriorates in old
queens (Butler 1957). In bumblebees, developing colonies reach a ‘competition point’ at which workers begin to reproduce, the timing of which is thought to depend on changes in queen-produced pheromone(s) (Alaux et al. 2007).

In the honeybee, a single queen-produced chemical (9-keto-2(E)-decenoic acid) induces near-complete worker sterility in bioassays (Kaatz et al. 1992); several other honeybee pheromones are known, but to our knowledge, there is little evidence that they directly affect worker sterility (although they may have an indirect effect via their positive chemotactic effect on workers; e.g. Hoover et al. 2003). Similarly, in termites, silencing one gene in the colony’s queen induces worker behaviour characteristic of recently de-queened colonies (Korb et al. 2009). Furthermore, the chemical profiles of reproductives frequently only differ from sterile workers by a single compound, or a single family of hydrocarbons (reviewed in Monnin 2006). In the present study, we showed that 3-MeC31, but not C31, affects worker sterility, even though both compounds are characteristic of queens; 3-MeC31 was also active in isolation. Based on these data, we suggest that worker sterility might often be regulated by single- rather than multi-component pheromones, and that at present there is insufficient evidence to rule out either of these hypotheses in any species. Single-component queen pheromones imply a mutualistic model of the origin of eusociality characterized by minimal parent–offspring conflict (Boomsma 2009), because in a high-conflict scenario, an evolutionary arms race over reproductive rights is predicted (Keller & Nonacs 1993). This arms race is expected to be characterized by the evolution of resistance to the queen pheromone in subordinates, followed by elaboration of the pheromone (e.g. by adding more component chemicals) and restoration of its manipulative effects (Le Conte & Hefetz 2008; Heinze & d’Ettorre 2009). Under the low-conflict model, workers might instead co-opt a single, arbitrary chemical that honestly indicates the presence of a healthy reproductive as a regulatory mechanism for their self-imposed sterility. Identification of queen primer pheromones in other taxa may reveal universal evolutionary trends and produce unexpected advances in our understanding of the origin and maintenance of eusociality.

We are grateful to all members of the Center for Social Evolution, Copenhagen, for a stimulating work environment, and to J. J. Boomsma for comments on the manuscript. This work was supported by the Marie Curie Excellence grant CODICES (MEXT-CT-2004-014202) assigned to P.d’E. and by a Marie Curie Intra-European Fellowship to L.H. (no. 235403; CHEMDOC).

REFERENCES


