Precise RNAi-mediated silencing of metabolically active proteins in the defence secretions of juvenile leaf beetles

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Allomones are widely used by insects to impede predation. Frequently these chemical stimuli are released from specialized glands. The larvae of Chrysomelina leaf beetles produce allomones in gland reservoirs into which the required precursors and also the enzymes are secreted from attached gland cells. Hence, the reservoirs can be considered as closed bio-reactors for producing defensive secretions. We used RNA interference (RNAi) to analyse in vivo functions of proteins in biosynthetic pathways occurring in insect secretions. After a salicyl alcohol oxidase was silenced in juveniles of the poplar leaf beetles, Chrysomela populi, the precursor salicyl alcohol increased to 98 per cent, while salicyl aldehyde was reduced to 2 per cent within 5 days. By analogy, we have silenced a novel protein annotated as a member of the juvenile hormone-binding protein superfamily in the juvenile defensive glands of the related mustard leaf beetle, Phaedon cochleariae. The protein is associated with the cyclization of 8-oxogranal to iridoids (methylcyclopentanoid monoterpenes) in the larval exudates made clear by the accumulation of the acyclic precursor 5 days after RNAi triggering. A similar cyclization reaction produces the secologandin part of indole alkaloids in plants.

Keywords: RNAi; insects; leaf beetle; secretome; salicyl alcohol oxidase; monoterpene cyclization

1. INTRODUCTION

Insects are extraordinarily inventive when it comes to producing defensive compounds for repelling their enemies. To circumvent auto-intoxicative effects, these natural products frequently originate in the epidermis-derived exocrine glands [1]. The gland cells produce secretions that are fortified with defensive compounds [2,3]. It has been demonstrated that insects convert either intrinsic precursors or food-derived compounds into biologically active allelochemicals [4–7]. The precursors can be activated in the defensive glands or in the secretions. Immature leaf beetles of the subtribe Chrysomelina, for example, produce their deterrents in biphasic secretions, and store them in nine unique pairs of impermeable reservoirs in their backs [8,9]. The larval exudates containing salicyl aldehyde (3) have been of particular interest [10,11]. The hydrophobic aldehyde forms an organic layer, accounting for 15 per cent of the total discharge volume, while the aqueous phase constitutes 85 per cent [12]. The latter contains the precursor salicyl alcohol (2) and a flavine-dependent salicyl alcohol oxidase (SAO); the SAO uses molecular oxygen as an electron acceptor for alcohol oxidation, yielding the aldehyde and hydrogen peroxide [12–14] (figure 1). Salicyl aldehyde is considered as a potent repellent against generalist predators [11,15] and as an antimicrobial agent [16]. The larvae feed on salicaceous plants and sequester the secondary metabolite salicin (1) [17–19]. After shuttling salicin to the defensive glands, the glucoside is cleaved by a β-glucosidase into 2 and glucose for further metabolism [20] (figure 1). According to phylogenetic analyses, the synthesis of 3 from sequestered precursors has evolved from the de novo production of defensive iridoids (methylcyclopentanoid monoterpenes containing an iridane skeleton) [21]. Also the last steps of the iridoid pathway in the secretions are thought to be similar to those found in sequestering species [20] (figure 1). At first, the sugar moiety is cleaved from 8-hydroxygeraniol-8-O-β-o-glucoside (4), and an oxygen-dependent oxidase converts the aglucone into 8-oxogranal (6) [20,22–24]. A subsequent cyclization reaction yields iridoids (7) [25]. Despite the many current genome- and transcriptome-sequencing projects, up to now it has only been shown for SAO sequences to be entangled in allomone production in the defensive secretions of the leaf beetle species Chrysomela tremulae, Chrysomela populi, Chrysomela lapponica and Phratora vitellinae [13,14,26]. To demonstrate the in vivo relevance of a target sequence, gene silencing by RNA interference (RNAi) is a suitable method. RNAi is an endogenous mechanism, derived from an anti-viral immune response [27], and can be found virtually in all eukaryotic species. It can be triggered artificially by double-stranded RNA (dsRNA), whose nucleotide sequence is identical to that of the target gene [28]. The

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RNAi effect is attended by decreased transcript and protein levels, and consequently by loss-of-function phenotypes. In addition to embryogenesis, pattern formation, reproduction and behaviour, RNAis allows biosynthetic pathways in insects to be successfully analysed [29–31].

Here, we describe how RNAi can be used to target the biosynthesis of discrete components in the defensive discharges of juvenile Chrysomelina. We first validated this technique by silencing the known enzyme(s) responsible for the biosynthesis of discrete components in the defensive secretions of C. populi. After knocking down the gene(s) encoding the enzyme(s), we showed that we are able to interrupt the biosynthesis in vivo. Next, we extended the method to the related de novo iridoid-producing species, P. cochleariae. In the secretions of its larvae 8-epi-loganin, one of the building blocks for more than 2500 indole alkaloids that have been isolated mainly from three plant families [32]. Although an enzyme with cyclase activity for seco-loganin biosynthesis has long been predicted, a corresponding sequence has yet to be published. In the P. cochleariae secretome, we identified a novel protein sequence in silico.

2. MATERIAL AND METHODS
See electronic supplementary material for complete secretome analyses by data-independent liquid chromatography/mass spectrometry detection (LC/MS<sup>n</sup>), cloning procedures, detailed quantitative real-time PCR procedure (qPCR), all primer sequences and accession numbers.

(a) Beetle rearing and secretion analyses
Chrysomela populi (L.) was collected near Dornburg, Germany (latitude 51.015, longitude 11.64), on Populus maximowiczii × Populus nigra. In the laboratory, beetles were kept in a 16 L : 8 D cycle, 18 ± 2 °C in light and 13 ± 2 °C in darkness. Phaedon cochleariae (F.) was laboratory-reared on Brassica oleracea cv. capitata var. alba (Gloria F1) in 16 L:8 D cycle conditions and 15 ± 2 °C. According to [33], we obtained the relative growth rate (RGR) of six biological replicates of each group of five larvae by RGR = [(final weight − weight of neonate larva)/(weight of neonate larva × developmental time (days))]. Each replicate group was weighed every 24 ± 3 h and data were compared with two-tailed <i>t</i>-test. Larval secretions were collected in glass capillaries (inner diameter, 0.28 mm; outer diameter, 0.78 mm, length 100 mm; Hirschmann, Eberstadt, Germany). Sealed capillaries containing samples were stored at −20 °C until needed. Secretions were weighed in the sealed capillaries on an ultra-microbalance (Metler-Toledo, Greifensee, Switzerland) three times; the weight of the capillaries was subtracted and the final weight was averaged.

(b) Production of double-stranded RNA
Sequenced plasmids pIB-CpopSAO (GeneBank: HQ245154.1) and pIB-PcTo-like (GeneBank: JQ728549) were used to amplify a 1.5 kb CpopSAO fragment and a 450 bp PcTo-like fragment, respectively. The gfp sequence was amplified from pcDNA3.1/CT-GFP-TOPO (Life Technologies, Darmstadt, Germany). The amplicons were subject to <i>in vitro</i> transcription assays according to instructions from the Ambion MEGAscript RNAi kit (Life Technologies, Darmstadt, Germany). The resulting dsRNA was eluted after nuclease digestion three times with 50 μl of injection buffer (3.5 mM Tris–HCl, 1 mM NaCl, 50 mM Na<sub>2</sub>HPO<sub>4</sub>, 20 mM KH<sub>2</sub>PO<sub>4</sub>, 3 mM KCl, 0.3 mM EDTA, pH 7.0). The concentration of dsRNA was calculated with 1 A<sub>260</sub> = 45 mg ml<sup>−1</sup> and adjusted to 1 μg μl<sup>−1</sup>. The quality of dsRNA was checked by TBE-agarose-electrophoresis.

(c) Injection of double-stranded RNA
First instar of C. populi with 5 mm body length was injected with 0.1–3 μg of dsRNA approximately 10 days after hatching. Phaedon cochleariae second instar with 4 mm body length was injected with 0.3 μg of dsRNA approximately 5 days after hatching. Injections were accomplished with ice-chilled larvae using a Nano2000 injector (WPI, Sarasota, FL, USA) directed by a three-axis micromanipulator. The larvae were injected parasagitally between the pro- and mesothorax.

(d) Off-target prediction
According to the mechanism of RNAi [28], the top and bottom strands of dsRNAs of CpopSAO, PcTo-like and gfp were diced in silico into all possible 21 bp fragments [34]. The resulting siRNAs were subjected to BLASTn
Guardian–column, 25 m. Substances were separated using Trace GC/MS 2000, Frankenhorst, Germany) equipped was subjected to GC/EIMS analysis (ThermoQuest Finnigan acetate and secretions of *P. cochleariae* were diluted in 1 : 150 (w/v) ethyl dichromethane. Of each diluted secretion, 1 μl was subjected to GC/EIMS analysis (ThermoQuest Finnigan Trace GC/MS 2000, Frankenhorst, Germany) equipped with Phenomenex (Aschaffenburg, Germany) ZB−5−W/ Guardian−column; column, 25 m. Substances were separated using helium as a carrier (1.5 ml min$^{-1}$). Conditions for *C. populi* secretions: 50°C (1 min), 10°C per minute to 80°C, 60°C per minute to 280°C (1 min). Inlet temperature was 220°C, transfer line was 280°C. Substances were identified according to standard substances 2 and 3. Conditions for *P. cochleariae* secretions: 50°C (2 min), 10°C per minute to 80°C, 5°C per minute to 200°C, 30°C per minute to 300°C (1 min). Inlet temperature was 220°C and transfer line was 280°C. Substances were identified according to [36] and the reference compounds 8-oxogeralial and chrysomelidial. The synthesis of 8-oxogeralial and chrysomelidial was carried out as in [25,37], respectively. Peak areas from GC-chromatograms were obtained using an ICIS-algorithm (*XCALIBUR BUNDLE* v. 2.0.7, Thermo Scientific).

### Statistical analyses
Two-tailed Student’s *t*-tests for unequal variation were used to value the significance levels of transcript abundances and to weight differences comparing values of three different biological replicates from the non-injected control (NIC) group with those of either the RNAi group or the *gfp* control. Multi-dimensional ANOVA tests were carried out to validate significant differences in time series and between different RNAi treatments. The level of significance was reached at a *p*-value of 0.05. Calculations were done with R (http://www.r-project.org/).

### 3. RESULTS

#### (a) Targeting the defensive glands of juvenile poplar leaf beetles by RNA interference
Recently, a 1872-bp *CpopSAO* cDNA (Genbank/ HQ245154.1) encoding a 69 kDa protein for conversion of 2 into 3 was identified from the larval defensive glands of *C. populi* ([13,14] (figure 2a)). It belongs to the glucose−methanol−choline (GMC) family of oxidoreductases [38]. Given that the expression of *CpopSAO* was detectable exclusively in glandular tissues (figure 2b), silencing this gene would affect only the process of glandular biosynthesis.

To induce RNAi in *C. populi* larvae, we injected 1.0 μg of 1.5 kb *CpopSAO* dsRNA into late first instar. A 719-bp dsRNA fragment of *gfp* served as a control for effects caused by dsRNA; although the RNAi machinery will be induced, genes should not be silenced. Furthermore, we included an NIC group in our experiments. By monitoring the developmental traits and the secretion production in *C. populi* and comparing the results with those from control groups, we found that silencing *CpopSAO* did not influence either growth rate or pupae weight (see the electronic supplementary material, figure S1a). But the larvae treated with *CpopSAO* dsRNA produced slightly more secretions than did the larvae of the control groups (see the electronic supplementary material, figure S1b), which might be owing to the different osmotic characteristics between 2 and 3 [12]. Because we did not detect significant differences between NIC and *gfp* controls in any experiments delineated below, we continue showing only the data of the *gfp* controls.

Transcript abundance was measured in glandular tissue using qPCR after 1, 3 and 12 days. Comparing tissue from these samples to tissue from the *gfp* controls, we noticed significant reductions to 7.6 per cent mRNA level (*p* = 0.002) just 24 h after injection. After day 3, the transcript level was diminished to 1.6 per cent (*p* = 0.004), and after day 12, to 0.5 per cent (figure 3a).

In accordance with the literature, SAO corresponds to the dominant band at 70 kDa in the secretions of *C. populi* and *P. cochleariae*.
The effects on the biosynthesis of 3 in the defensive secretions were determined by GC/MS analysis. For these experiments, 0.1, 1.0 and 3.0 µg of CpopSAO dsRNA were injected into larvae from the same clutch. As in the protein reduction, we detected 2 in the defensive secretions just 2 days after the injection of 3.0 µg CpopSAO dsRNA (figure 3c). Compound 2 was not detectable in gfp control secretions. In addition, no unexpected chemical compound arose owing to the dsRNA treatments. By setting the peak area of 3 in ratio equalling the sum of the main peak areas, a diagram of the RNAi-dependent reduction of 3 can be plotted (figure 3d). We have tested dsRNA amounts ranging from 0.1 to 3.0 µg. After RNAi induction, significantly less aldehyde was observed for the 3.0 µg CpopSAO group (p = 0.015) on the 4th day and for all tested CpopSAO groups on the 5th day (0.1 µg, p = 0.016; 1 µg, p = 0.002; 3 µg, p = <0.001). Biological variation prevented us from observing dose-dependent RNAi effects in these experiments; the amount of 2 did not differ significantly between the RNAi samples.

(b) Off-target prediction and validation for CpopSAO
Owing to strong sequence identities, co-silencing non-target genes can cause unintended side-effects [39,40]. Therefore, we performed off-target predictions for the desired dsRNA sequences of CpopSAO and gfp. Predicted off-target genes were validated by qPCR using cDNA derived from successful RNAi experiments. Because of a lack of genome sequences, the potential silencing effects of targeting the nucleus where fragments of the long dsRNA may bind to non-transcribed regulatory sequences [41] or introns [42] could be neither predicted nor validated.

For gfp dsRNA, no critical candidates were detected in the transcriptome library of C. populi. Off-target analyses in the C. populi sequence library, however, identified 25–21 bp contiguous regions of CpopSAO dsRNA that were identical to sequences of eight unique transcripts (see the electronic supplementary material, table ST2 for putative off-target hits). Three of them encode putative proteins having the GMC-oxidoreductase motif in the C-terminal region (CpGMClike-I-III) and five were annotated as hypothetical proteins (CpCOMP3092; CpCOMP6024; CpCOMP36289; CpCOMP38777; CpCOMP51471).

CpopSAO shares with CpGMClike-I two similar regions spanning 22 and 25 nucleotides (nts) each; these regions are interrupted by one mismatch (22/1 and 25/1) and, with CpGMClike-III, one similar sequence stretch without mismatch (24/0). CpGMClike-II and the five remaining transcripts possess sequence regions of 22/0 to 20/0 nts identical to CpopSAO. In all tissues, all putative off-target genes exhibited generally low expression levels with relative Cq values median less than 2 × 10⁻³. qPCR assays were carried out for all eight targets 12 days after larvae were treated with 1.0 µg 1500-bp CpopSAO dsRNA; only for the CpGMClike-I and CpGMClike-II did these treatments reveal significant differences of transcript level in the gut tissue (p = 0.049; p = 0.032). No other tested transcripts showed changes of mRNA abundance in the examined tissues.
(see the electronic supplementary material, figure S2). Since *C. populi* larvae transport the plant-derived precursor into the defensive glands for final transformation, we assume that the off-target effects on the putative GMC-oxidoreductases in gut tissue of unknown function do not distort the RNAi effects observed in the secretions.

**c) Identification of unknown proteins in the defence-related secretome of Phaedon cochleariae**

After successfully introducing the ‘lack-of-function approach’ to the defensive secretions of *C. populi* by silencing an enzyme for which we had a clear expectation of the resulting phenotype, we used the method to identify proteins in unknown secretions. For this reason, we chose the larval exudates from the related de novo iridoid-producing species *P. cochleariae*. We assigned to the abundant 35-kDa band a putative protein whose deduced sequence contains 243 amino acids and a 22 amino acid signal peptide; the existence of such a sequence suggests that the mature protein is secreted (figure 2a). It possesses a conserved domain (pfam06585) characteristic for the juvenile hormone-binding protein (JHBP) superfamily. Sequence comparisons using the BLAST algorithm [35] revealed that the *P. cochleariae* amino acid sequence shares only very limited identity with functionally characterized insect proteins, for example, 12 per cent identity with the JHBP from *Bombyx mori* [43] and 16 per cent with the takeout (To) 1 from *Epiphyas postvittana* [44] (figure 2a). Higher identities up to 25 per cent were found only with insect proteins not yet fully characterized in their functions, such as those with the To-like protein (XP_001191952) from *Acyrthosiphon pisum* or the JHBP-like (XP_001359416) from *Drosophila pseudoobscura pseudoobscura*. None of the mentioned insect species is known to produce cyclic monoterpenoids.

The JHBP superfamily combines both the To protein family and the JHBP family. There are two major differences between the families: one is the number of disulphide bonds (To proteins have one and JHBPs have two) and the second are the conserved C-terminal sequence motives that are only present in To proteins. *In silico* analyses predicted in the *P. cochleariae* sequence seven N-glycosylation sites and only one disulphide bond. Along with identifying the two To-specific motives [45] (figure 4) in the C-terminal region, we conclude that our protein can be attributed to the To family. Therefore, we named it *Pc*To-like.

Despite the generally low sequence similarity, most To proteins and JHBPs are classified as ligand-binding proteins for juvenile hormones or similar hydrophobic terpenoids [44,46–49]. Because the precursor of the cyclic iridoid is also a terpenoid, we hypothesize that the putative protein could be involved in the iridoid biosynthesis in the defensive secretions. The assumption that the putative protein has relevance in the defensive glands is corroborated by the high transcript level which has been detected mainly in the glandular tissue of juvenile mustard leaf beetles (figure 2c). Low mRNA levels were also detectable in the fat body tissue.

**d) RNA interference effects in larvae of Phaedon cochleariae**

A total of 0.3 μg of dsRNA derived from a 450-bp fragment of the *Pc*To-like sequence was injected into second instars of *P. cochleariae*. Transcript quantification 5 days after dsRNA injection confirmed a significant reduction of the mRNA in glandular tissue (p = 0.043) down to 1.0 per cent (+0.9%) compared with mRNA levels in *gfp* injections (figure 5a).

Phenotypic analyses after injection of dsRNA on the composition of low-molecular-weight compounds in the secretions were carried out by using GC/MS. The quality of the metabolites in samples collected 1 and 2 days after *Pc*-To-like RNAi induction did not vary from the quality of the metabolites in those collected from *gfp* controls. In both treatments, we detected only the end-product 7. The first deviation in the composition of the secretions was measured 3 days after dsRNA injection. Only in samples triggered by *Pc*-To-like RNAi did minor amounts of the postulated intermediate 6 in addition to 7 emerge. After 5 days, however, 6 clearly accumulated in addition to 7 owing to the RNAi effect (figure 4b). Therefore, we conclude that the *Pc*-To-like has to be involved in the cyclization of monoterpenoid precursors into iridoids.

Off-target effects were predicted using the method described for *CpopSAO*, and predicted off-target effects were avoided from final dsRNA sequence by choosing the template for dsRNA outside of areas of predicted off-target effects.

**4. DISCUSSION**

The results of our larval RNAi experiments clearly demonstrate selective excision of a component in a biosynthetic pathway. To the best of our knowledge, RNAi has never been used to target enzymes in insect defensive secretions. Owing to the silencing of *CpopSAO*, the chemical composition in the larval exudates of *C. populi* was massively altered, starting as early as 48 h after treatment. This shows a distinct function of this enzyme in vivo. Before, Kirsch et al. [13] showed activity only in *in vitro* assays. Evidently, RNAi is a valuable technique for identifying in vivo relevance for unknown proteins in defensive glands. Although insects contain a large number of exocrine glands in which bioactive compounds are produced, to date few studies have relied on RNAi to provide evidence for the in vivo function of enzymes in insect glands. One example is the production of sex pheromones in special glands of the silkworm *Bombyx mori*. By injecting the pupae with dsRNA, Ohnishi et al. [50,51] were able to dissect the components of the biosynthetic pathway as well as assign a function to a transport protein within the glands of adults. Another RNAi target was the production of pheromone in jewel wasps, *Nasonia vitripennis*. Silencing an epoxide hydrolyase in these insects resulted in pheromone reduction by 55 per cent and suppressed the targeted gene transcripts by 95 per cent [52]. Freshly emerged males were injected and 2 days later levels of transcript and pheromones were analysed. As our results demonstrate, RNAi effects are easily detectable in exocrine glands. In the secretions of immature *P. cochleariae*, we were able to assign in vivo relevance to a cDNA encoding a protein which is important for the cyclization of iridoids. The iridoid pathway in insects was already proposed by using deuterium labelled precursors by Weibol et al. [53]. In his work, the stereospecificity of the cyclization was analysed and allocated to an enzymatic conversion. However, to date JHBPs and
larval defence secretions. Functional enzymes in Chrysomelina and clarify the molecular machinery for the biosynthesis of deterrents in on-going experiments will reveal more. Therefore, exudates has to be analysed binding in To proteins [44]. Most of the putative To proteins have been established as being carriers of hydrophobic ligands [44,48]. Several lines of evidence indicate that JHBPs form complexes with juvenile hormones (JHs) which provide protection of the chemically labile JHs against nonspecific enzymatic degradation and/or adsorption to lipophilic surfaces during the delivery process from the production site to the target tissue [46,47,49]. Up to now only the crystal structure of To 1 from B. mori has been elucidated [50,51]. Although the specificity. Off-target effects can arise when siRNAs are available [34]. These approaches are not suitable for organisms whose genomes have yet to be sequenced. In the last few years, several approaches have been used to detect off targets for those species, such as screening for ing degradation of similar sequences [39]. For sequenced sequence similarity to non-target mRNA and thus trigger- dicated from long dsRNA fragments possess sufficient specificity. Off-target effects can arise when siRNAs To perform RNAi experiments, it is essential to ensure the specificity. Off-target effects can arise when siRNAs duced from long dsRNA fragments possess sufficient sequence similarity to non-target mRNA and thus triggering degradation of similar sequences [39]. For sequenced organisms, genome-scale off-target prediction programs are available [34]. These approaches are not suitable for organisms whose genomes have yet to be sequenced. In the last few years, several approaches have been used to detect off targets for those species, such as screening for ing degradation of similar sequences [39]. For sequenced sequence similarity to non-target mRNA and thus trigger-

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Figure 4. Amino acid alignments (ClustalW) of Pc-To-like from P. coehlaeri and other members from the To/JHBP family (Epispyias postvittana To1: GeneBank: ACDF39401; Acyrthosiphon pisum To-like: GeneBank: XP_001952685; Phorinia regina To-like protein: GeneBank: BAD83405; Bombyx mori JHBP: GeneBank: AAF19267). Solid black shading depicts amino acids identical to Pc-To-like sequence. White inverted triangles indicate the predicted signal peptide of Pc-To-like. Asterisks mark the predicted N-glycosylation sites. Conserved cysteine residues that form disulphide bonds are marked with a bracket above the sequence. The two black boxes show the location of the To-specific motifs.

To proteins have been established as being carriers of hydrophobic ligands [44,48]. Several lines of evidence indicate that JHBPs form complexes with juvenile hormones (JHs) which provide protection of the chemically labile JHs against nonspecific enzymatic degradation and/or adsorption to lipophilic surfaces during the delivery process from the production site to the target tissue [46,47,49]. Up to now only the crystal structure of To 1 from E. postvittana with ubiquinone provided direct evidence for ligand binding in To proteins [44]. Most of the putative To proteins await elucidation of their mode of action. Therefore, the actual mechanism how Pt-To-like acts in the defensive exudates has to be analysed in vitro with purified recombinant protein. On-going experiments will reveal more functional enzymes in Chrysomelina and clarify the molecular machinery for the biosynthesis of deterrents in larval defence secretions.

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dsRNA resulted in the production of 

\[ 100\% = \Delta Cq \text{ of } gfp\text{-control} \]

transcript abundance (%)

Asterisks indicate level of significance: *p < 0.05.

(b) GC-chromatogram of diluted secretions on day 5 after treatment, above: injecting 0.3 \( \mu g \) \( PcTo-like \) dsRNA resulted in the production of 6 and 7; below: injecting 0.3 \( \mu g \) \( gfp \) dsRNA resulted in the production of 7.

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R.R.B., P.R., M.S., M.K., N.W., W.B. and A.B. designed study; R.R.B. established RNAi in leaf beetles and performed RNAi treatments of \( C. populi \) and its control treatments, performed off-target validation, collected all corresponding data except the off-target prediction and analysed output data; P.R. identified \( PcTo-like \), performed RNAi treatments of \( PcTo-like \) and control treatments, collected all corresponding data and analysed output data. S.F., M.S. and M.G. generated transcriptome libraries. M.S. established and performed off-target prediction and contributed to interpretation of LC/MS\( ^2 \) output data. M.K. designed GC/MS assays, synthesized 6 and 7 and contributed to interpretation of output data. N.W. performed LC/MS\( ^2 \) analysis, collected and contributed to interpretation of output data. W.B. and A.B. contributed substantially to interpretation of all output data. R.R.B., P.R. and A.B. wrote first draft of the manuscript, and all authors contributed substantially to revisions.

**REFERENCES**


