Honest sexual signalling mediated by parasite and testosterone effects on oxidative balance

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Extravagant ornaments evolved to advertise their bearers’ quality, the honesty of the signal being ensured by the cost paid to produce or maintain it. The oxidation handicap hypothesis (OHH) proposes that the main cost of testosterone-dependent ornamentation is oxidative stress, a condition whereby the production of reactive oxygen and nitrogen species (ROS/RNS) overwhelms the capacity of antioxidant defences. ROS/RNS are unstable, very reactive by-products of normal metabolic processes that can cause extensive damage to key biomolecules (cellular proteins, lipids and DNA). Oxidative stress has been implicated in the aetiology of many diseases and could link ornamentation and genetic variation in fitness-related traits. We tested the OHH in a free-living bird, the red grouse. We show that elevated testosterone enhanced ornamentation and increased circulating antioxidant levels, but caused oxidative damage. Males with smaller ornaments suffered more oxidative damage than those with larger ornaments when forced to increase testosterone levels, consistent with a handicap mechanism. Parasites depleted antioxidant defences, causing oxidative damage and reduced ornament expression. Oxidative damage extent and the ability of males to increase antioxidant defences also explained the impacts of testosterone and parasites on ornamentation within treatment groups. Because oxidative stress is intimately linked to immune function, parasite resistance and fitness, it provides a reliable currency in the trade-off between individual health and ornamentation. The costs induced by oxidative stress can apply to a wide range of signals, which are testosterone-dependent or coloured by pigments with antioxidant properties.

Keywords: oxidative stress; antioxidant; ornament; trade-off; red grouse Lagopus lagopus scoticus; Trichostrongylus tenius

1. INTRODUCTION

Testosterone plays a pivotal role in regulating the expression of many animal ornaments (Folstad & Karter 1992; Wingfield et al. 2001). Males typically benefit from elevated testosterone levels and enhanced ornamentation in intra- and inter-sexual contexts, but maintaining high testosterone levels may be costly (Folstad & Karter 1992). A main cost might be a reduced ability to resist parasites: according to the immunocompetence handicap hypothesis (ICHH) testosterone impairs immune function, so only individuals of high genetic quality can endure the cost of displaying larger ornaments (Folstad & Karter 1992). The idea that testosterone may suppress the immune system in birds has, however, received mixed support (Roberts et al. 2004), possibly because the costs are not entirely mediated through the physiological pathways hitherto examined (Owen-Ashley et al. 2004; Mougeot et al. 2005b; Blas et al. 2006). The oxidation handicap hypothesis (OHH), a refinement of the ICHH, proposes that the trade-off ensuring honest signalling is between ornamentation and oxidative stress (von Schantz et al. 1999; Alonso-Alvarez et al. 2007). Elevated testosterone can lead to increased reactive oxygen and nitrogen species (ROS/RNS) production and oxidative stress (Alonso-Alvarez et al. 2007), which may, in turn, impair lymphocyte proliferation and signalling pathways involved in an immune response (Larbi et al. 2007). Additionally, immune system activation produces ROS/RNS to help counter invading pathogens (Romero et al. 1998; Hörak et al. 2007), but their overproduction can lead to oxidative stress, incurring damage to host tissues including ornaments, particularly when individuals lack sufficient antioxidant protection (von Schantz et al. 1999; Splettstoesser & Schull-Werner 2002; Halliwell & Gutteridge 2007). Testosterone-dependent ornaments may thus be inherently vulnerable to oxidative stress, itself intimately linked to immune function and parasite resistance (von Schantz et al. 1999). The OHH therefore proposes that only high-quality individuals, with a prime antioxidant system, could afford the costs (increased oxidative stress) of maintaining high testosterone levels and enhanced ornamentation.

We tested the OHH in free-living red grouse (Lagopus lagopus scoticus). This bird displays supra-orbital red combs, the coloration of which is carotenoid based (Mougeot et al. 2007a,b) and whose size is testosterone dependent (Mougeot et al. 2005a). Comb size functions

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in intra- and inter-sexual selection: males with higher testosterone levels and bigger combs benefit by being dominant, more aggressive, holding larger territories and being more attractive to females (Mougeot et al. 2003; Redpath et al. 2006). Bigger combs are often, but not always, redder, and seem likely to contain absolutely more carotenoids although this remains to be tested. Therefore, the possibility exists that the comb size and comb colour reveal similar information about individual quality. However, the signalling function of comb colour remains untested, so we focus our study on comb size. Using a factorial experimental design, we manipulated (i) parasite burdens (P) of the nematode Trichostrongylus tenuis, which has well-known negative effects on this host (Hudson 1986; Delahay et al. 1995), using experimental infections (Hudson 1986; Delahay et al. 1995; Mougeot et al. 2005b) and (ii) testosterone levels (T) using implants (Mougeot et al. 2005a). We initially purged males of T. tenuis (Hudson 1986) and began manipulations 15 days later.

We then randomly assigned males to one of four treatment groups (10 in each): (i) empty implants, no parasite challenge (T−P− males; control group), (ii) empty implants, challenge with T. tenuis infective larvae (T−P+ males), (iii) testosterone implants, no parasite challenge (T+P− males), and (iv) testosterone implants, challenge with T. tenuis (T+P+ males). We sampled males before treatments (S1) and again after 10 days (S2) and 17 days (S3; see appendix A). We investigated treatment effects on (i) plasma testosterone concentration, (ii) T. tenuis abundance, (iii) ornamentation (comb area), (iv) total antioxidant status (TAS), (v) circulating antioxidant defences, and (vi) plasma concentrations of malondialdehyde (MDA), a measure of oxidative damage. TAS measures the capacity of the plasma to quench a free radical cation and the pooled effect of all extracellular, non-enzymatic antioxidants in plasma (e.g. uric acid, vitamins C and E, carotenoids) (von Schantz et al. 1999; Halliwell & Gutteridge 2007). MDA is formed when lipid hydroperoxides break down, a process (lipid peroxidation) caused by oxidative stress (Romero et al. 1998).

We predicted that (i) increased testosterone levels would enhance ornamentation but increase oxidative damage, measured in terms of MDA, (ii) a developing parasite infection would reduce circulating antioxidants, cause oxidative damage and reduced ornamentation, (iii) the cost of each treatment (oxidative damage) would depend on initial ornament size, with males displaying smaller combs showing a greater increase in oxidative damage (MDA) relative to larger combed males, and (iv) the treatment effects on oxidative damage and the ability of males to increase antioxidant activity (TAS) to reduce this damage would explain changes in male ornamentation.

2. MATERIAL AND METHODS

(a) Experiment

We worked on Edinglassie and Catterick moors (UK) in 2006. In September, we captured 20 male red grouse on each site, by dazzling and netting them at night (Hudson 1986). Upon first capture (S0; see appendix A), we fitted males a radio-collar (TW3-necklace radio-tags, Biotrack) and gave each a 1 ml of levamisole hydrochloride (Milverm Gold, Schering-Plough Animal Health, Welwyn Garden City, UK) to purge them of their T. tenuis nematodes (Hudson 1986).

We started the experiment 15 days later, allowing birds enough time to clear the anthelminthic. At S1, we gave males hormone and parasite treatments (five males per treatment per site). Males were implanted with two silastic tubes (each 20 mm long, 1.57 mm inner and 2.41 mm outer diameter) sealed with glue at both ends. T− males were given two empty implants, and T+ males two implants filled with crystalline testosterone propionate (Sigma Aldrich, UK) to elevate testosterone for two to three months (Mougeot et al. 2003). Implants were inserted subcutaneously on the flank following local anaesthesia. P+ males received an oral dose of water containing approximately 5000 T. tenuis infective larvae and P− males only water. We sampled males upon treatment (S1), 10 days later (S2) and 17 days later (S3). Details on the timing and data sampling for the experiment are given in appendix A. We held all the necessary Home Office licences for conducting the procedures described in this work (Licence PPL80/1437).

(b) Measurements and blood sampling

We measured comb area (maximum length × width of flattened comb) as an index of ornament size (Mougeot et al. 2005a). We took a blood sample from the brachial vein, separated plasma by centrifugation (2 min at 7000 rpm) and froze the samples in liquid nitrogen within 5 min of collection. Plasma samples were taken to the laboratory afterwards and stored at −80°C.

(c) Testosterone assays

Plasma testosterone concentration was measured using a commercially available testosterone enzyme immunoassay (Elisa Kit EIA-1559 from DRG Diagnostics, Marburg, Germany), an assay that has been developed and validated for determining testosterone levels in small volume (20 μl) avian plasma samples (Washburn et al. 2007). Intra- and inter-assay coefficients of variation were 3.59 and 7.14 per cent, respectively, and the detection limit was 0.2 ng ml−1. Repeatability was determined on a subsample measured twice (r = 0.88; n = 30, p < 0.001).

(d) Lipid peroxidation assays

Plasma concentrations of MDA were calculated by HPLC using fluorescence detection (Agarwal & Chase 2002). All chemicals were HPLC grade, and chemical solutions were prepared using ultra pure water (Milli-Q Synthesis; Millipore, Watford, UK). Assays were carried out in 2 ml capacity screw-top microcentrifuge tubes. To a 15 μl aliquot of sample or standard (1,1,3,3-tetraethoxypropane, TEP; see below), 15 μl butylated hydroxytoluene solution (0.05% w/w in 95% ethanol), 120 μl phosphoric acid solution (0.44 M) and 30 μl thiobarbituric acid solution (42 mM) were added. Samples were capped, vortex mixed for 5 s, then heated at 100°C for 1 h on a dry bath incubator to allow formation of MDA-TBA adducts. Samples then cooled on ice for 5 min, before 75 μl n-butanol was added and tubes were vortex mixed for 30 s. Tubes were then centrifuged at 12 000 × g for 1 h on a dry bath incubator to allow formation of MDA-TBA adducts. Samples were then cooled on ice for 5 min, before 75 μl n-butanol was added and tubes were vortex mixed for 30 s. Tubes were then centrifuged at 12 000 × g for 1 h on a dry bath incubator to allow formation of MDA-TBA adducts. Samples were then heated at 100°C for 1 h on a dry bath incubator to allow formation of MDA-TBA adducts. Samples were then cooled on ice for 5 min, before 75 μl n-butanol was added and tubes were vortex mixed for 30 s. Tubes were then centrifuged at 12 000 × g for 1 h on a dry bath incubator to allow formation of MDA-TBA adducts. Samples were then cooled on ice for 5 min, before 75 μl n-butanol was added and tubes were vortex mixed for 30 s. Tubes were then centrifuged at 12 000 × g for 1 h on a dry bath incubator to allow formation of MDA-TBA adducts. Samples were then cooled on ice for 5 min, before 75 μl n-butanol was added and tubes were vortex mixed for 30 s. Tubes were then centrifuged at 12 000 × g for 1 h on a dry bath incubator to allow formation of MDA-TBA adducts.

Proc. R. Soc. B (2009)
sulphonate). Hydrogen peroxide (H2O2) was then added and metmyoglobin and ABTS (2,2-azino-di-[3-ethylbenzthiazoline-6-sulfonic acid]) were incubated for 15 s with a chromogen composed of ABTS, which generates a blue-green colour. Colour is measured at 600 nm before and after H2O2 addition, thus indicating that the initial parasite purging had been effective. Testosterone levels increased more in T+ than in T− males between S1 and S2 (table 1; figure 1a), but increased T. tenuis abundance. When challenged (S1), males had no detectable T. tenuis worms. However, by S3, average T. tenuis abundance was higher in P+ (mean 147 worms) than in P− males (15 worms), irrespective of the testosterone treatment (figure 1b; P(Treat): F1,25 = 10.57, p = 0.001; TTreat: F1,25 = 10.46, p = 0.049; P(Treat)×TTreat: F1,25 = 0.10, p = 0.74).

Consistent with prediction 1, testosterone implants increased comb area and the concentration of MDA. Comb area increased more in T+ than in T− males between S1 and S2 and remained larger afterwards (table 1; figure 1c). MDA concentration increased more in T+ than in T− males (table 1; figure 1d).

Consistent with prediction 2, parasite challenge caused oxidative damage (MDA concentration increased more in P+ than in P− males; table 1; figure 1d). MDA concentration increased more in T+ than in T− males while it increased in T−P+ males between S2 and S3; table 1; figure 1c).

Consistent with prediction 3, changes in MDA were dependent on initial comb area, but in T+ males only. In T+ males, changes in MDA were not explained by parasite treatment, but by initial comb area (P(Treat): F1,14 = 0.77, p = 0.39; comb: F1,14 = 4.68, p < 0.05; slope: −0.004 ± 0.002). In T− males, changes in MDA

3. RESULTS

At the start of the experiment (S1), males that had been dosed previously with anthelmintic (at S0) had no T. tenuis eggs in their faeces (n = 5, 4, 7 and 10 males sampled in the T−P−, T+P−, T−P+ and T+P+, respectively), indicating that the initial parasite purging had been effective. Testosterone levels increased more in T+ than in T− males between S1 and S2 (table 1; figure 1a) and remained higher afterwards (S2 until S3). Parasite challenges had no effect on testosterone levels (table 1; figure 1a), but increased T. tenuis abundance. When challenged (S1), males had no detectable T. tenuis worms. However, by S3, average T. tenuis abundance was higher in P+ (mean 147 worms) than in P− males (15 worms), irrespective of the testosterone treatment (figure 1b; P(Treat): F1,25 = 10.57, p = 0.001; TTreat: F1,25 = 10.46, p = 0.049; P(Treat)×TTreat: F1,25 = 0.10, p = 0.74).

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(e) Total antioxidant status

TAS concentration of plasma was assessed by means of commercial kits (Randox Laboratories Ltd, Crumlin, UK) adapted to an automated spectrophotometer (A25- Autoanalyzer; Biosys S.A., Barcelona). Plasma samples were incubated for 15 s with a chromogen composed of metmyoglobin and ABTS (2,2-azino-di-[3-ethylbenzthiazoline-6-sulfonate]). Hydrogen peroxide (H2O2) was then added and the sample was incubated for 195 s. Hydrogen peroxide (H2O2) addition induces the production of the radical cation ABTS, which generates a blue-green colour. Colour is measured at 600 nm before and after H2O2 addition, thus determining the change in colour. Antioxidants in the plasma sample cause suppression of this colour change to a degree that is proportional to their concentration. Results are given as mmol l−1 of plasma. Repeatability was determined on a subsample measured twice (r = 0.92, n = 30, p < 0.001).

(f) Parasite counts, cultures and challenges

We estimated T. tenuis abundance using either caecal egg counts (from caecal samples collected from captured males at S1) or direct worm counts (from caeca collected from males humanly killed at the end of the experiment, at S3). Caecal egg counts provide reliable estimates of worm burdens and were used to calculate T. tenuis abundance (Seivwright et al. 2004). More details regarding the methods for estimating T. tenuis abundance from caecal egg counts or direct worm counts, and for cultivating larvae for challenges are given elsewhere (Shaw 1988; Moss et al. 1990; Seivwright et al. 2004; Mougeot et al. 2005b).

(g) Statistical analyses

We used SAS v. 8.01 (SAS 2001). Counts of T. tenuis worms were fitted to generalized linear models using a Poisson error distribution. We calculated individual changes over time in study parameters (testosterone, comb area, MDA and TAS) as the difference between the final and initial values corrected for initial values. MDA and TAS were measured at S1 and S2 only.

### Table 1. Effect of testosterone and parasite treatments on changes over time in testosterone concentration (ΔTtestosterone), ornamentation (Δcomb area), plasma concentration of malondialdehyde (ΔMDA) and total antioxidant status (ΔTAS).

<table>
<thead>
<tr>
<th>Treatment effects:</th>
<th>Sampling time</th>
<th>d.f.</th>
<th>F (P-value)</th>
<th>F (P-value)</th>
<th>F (P-value)</th>
</tr>
</thead>
<tbody>
<tr>
<td>ΔTtestosterone</td>
<td>S1–S2</td>
<td>1,22</td>
<td>17.76 (&lt;0.001)</td>
<td>0.96 (0.35)</td>
<td>0.92 (0.35)</td>
</tr>
<tr>
<td>ΔTtestosterone</td>
<td>S2–S3</td>
<td>1,11</td>
<td>3.59 (0.08)</td>
<td>0.75 (0.47)</td>
<td>0.56 (0.47)</td>
</tr>
<tr>
<td>Δcomb area</td>
<td>S1–S2</td>
<td>1,32</td>
<td>80.01 (&lt;0.001)</td>
<td>0.61 (0.16)</td>
<td>0.7 (0.47)</td>
</tr>
<tr>
<td>Δcomb area</td>
<td>S2–S3</td>
<td>1,30</td>
<td>0.95 (0.34)</td>
<td>1.91 (0.17)</td>
<td>2.7 (&lt;0.05)</td>
</tr>
<tr>
<td>ΔMDA</td>
<td>S1–S2</td>
<td>1,28</td>
<td>5.32 (&lt;0.05)</td>
<td>10.88 (&lt;0.01)</td>
<td>0.01 (0.92)</td>
</tr>
<tr>
<td>ΔTAS</td>
<td>S1–S2</td>
<td>1,31</td>
<td>16.1 (&lt;0.001)</td>
<td>4.31 (&lt;0.05)</td>
<td>3.41 (0.07)</td>
</tr>
</tbody>
</table>

Note: ΔTtestosterone = testosterone treatment; T− males, sham implanted; T+ males, implanted with testosterone.

### Results

At the start of the experiment (S1), males that had been dosed previously with anthelmintic (at S0) had no T. tenuis eggs in their faeces (n = 5, 4, 7 and 10 males sampled in the T−P−, T+P−, T−P+ and T+P+, respectively), indicating that the initial parasite purging had been effective. Testosterone levels increased more in T+ than in T− males between S1 and S2 (table 1; figure 1a) and remained higher afterwards (S2 until S3). Parasite challenges had no effect on testosterone levels (table 1; figure 1a), but increased T. tenuis abundance. When challenged (S1), males had no detectable T. tenuis worms. However, by S3, average T. tenuis abundance was higher in P+ (mean 147 worms) than in P− males (15 worms), irrespective of the testosterone treatment (figure 1b; P(Treat): F1,25 = 10.57, p = 0.001; TTreat: F1,25 = 10.46, p = 0.049; P(Treat)×TTreat: F1,25 = 0.10, p = 0.74).

Consistent with prediction 1, testosterone implants increased comb area and the concentration of MDA. Comb area increased more in T+ than in T− males between S1 and S2 and remained larger afterwards (table 1; figure 1c). MDA concentration increased more in T+ than in T− males (table 1; figure 1d).

Consistent with prediction 2, parasite challenge caused oxidative damage (MDA concentration increased more in P+ than in P− males; table 1; figure 1d). MDA concentration increased more in T+ than in T− males while it increased in T−P+ males between S2 and S3; table 1; figure 1c).

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were explained by parasite treatment, irrespective of initial comb area (GLM: P treat: F 1,13 = 7.36, p < 0.05; comb: F 1,13 = 0.90, p = 0.23; slope ± s.d.: +0.002 ± 0.002).

TAS increased more in T+ than in T− males, while parasite treatment reduced TAS, but depending on testosterone treatment (Table 1). In T+ males, elevated testosterone caused an increase in the levels of circulating antioxidant defences, irrespective of parasite treatment (PTreat: F 1,14 = 0.11, p = 0.74; Figure 1c). In T− males, parasite treatment reduced TAS (F 1,14 = 8.09; p < 0.05; Figure 1e).

Before testosterone and parasite treatments (at S1), MDA was not significantly related to TAS (F 1,26 = 1.22, p = 0.28; slope ± s.d.: +0.404 ± 0.365). However, changes in TAS and MDA explained the impact of treatments on changes in comb area after taking into account treatment group level effects (Table 2). Changes in MDA explained the extent to which testosterone implants initially increased ornamentation (between S1 and S2), depending on testosterone treatment (Table 2; significant ΔMDA × TTreat interaction). In T+ males, individuals that increased their comb area most were least susceptible to oxidative stress (F 1,11 = 10.09, p < 0.01; Figure 2b), while no such effect was found in T− males (F 1,10 = 0.24, p = 0.63; Figure 1a).

Consistent with prediction 4, changes in TAS explained lagged changes in ornamentation, between S2 and S3 (Table 2; Figure 2c,d). In T− males, a reduction in
TAS was associated with a decrease in ornament size (figure 2c). In T− males, a greater increase in TAS was associated with a continued increase in ornamentation (figure 2d).

4. DISCUSSION

Testosterone treatment successfully increased testosterone levels, which were higher in T− than in T+ males after implantation (S2 and S3), but were still within the natural range (Mougeot et al. 2005a). Parasite treatment also successfully increased T. tenuis infection levels (higher in P+ than in P− males at S3). There was no short-term effect of testosterone on the effectiveness of parasite challenges, although previous work showed that elevated testosterone can indirectly increase T. tenuis abundance 1 year after challenge (Mougeot et al. 2005b; Seivwright et al. 2005).

Testosterone implants enhanced ornamentation (prediction 1), while parasite challenges reduced ornamentation (prediction 2), but only in T− males, and with a time lag. Trichostrongylus tenuis larvae impact most

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**Table 2. Effect of treatments, changes in plasma concentration of malondialdehyde (ΔMDA) and antioxidant activity (ΔTAS) on changes in ornamentation (Δcomb) at different sampling times (S1–S2; S2–S3).**

<table>
<thead>
<tr>
<th>Dependent variable</th>
<th>d.f.</th>
<th>Δcomb S1–S2</th>
<th>F</th>
<th>p-value</th>
<th>Δcomb S2–S3</th>
<th>F</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>T Treat^b^</td>
<td>1,19</td>
<td>84.12</td>
<td>&lt;0.001</td>
<td>0</td>
<td>0.96</td>
<td>0.05</td>
<td>0.01</td>
</tr>
<tr>
<td>P Treat^c^</td>
<td>1,19</td>
<td>1.09</td>
<td>0.31</td>
<td>4.98</td>
<td>&lt;0.05</td>
<td>0.12</td>
<td>0.37</td>
</tr>
<tr>
<td>T Treat×P Treat</td>
<td>1,19</td>
<td>0.06</td>
<td>0.8</td>
<td>3.19</td>
<td>0.09</td>
<td>0.32</td>
<td>0.27</td>
</tr>
<tr>
<td>ΔMDA</td>
<td>1,19</td>
<td>13.07</td>
<td>&lt;0.01</td>
<td>1.29</td>
<td>0.27</td>
<td>0.17</td>
<td>0.3</td>
</tr>
<tr>
<td>ΔTAS^d^</td>
<td>1,19</td>
<td>0.46</td>
<td>0.51</td>
<td>8.54</td>
<td>&lt;0.01</td>
<td>0.05</td>
<td>0.99</td>
</tr>
<tr>
<td>ΔMDA×T Treat</td>
<td>1,19</td>
<td>4.09</td>
<td>&lt;0.05</td>
<td>0.82</td>
<td>0.37</td>
<td>0.17</td>
<td>0.3</td>
</tr>
<tr>
<td>ΔTAS×P Treat</td>
<td>1,19</td>
<td>0.39</td>
<td>0.54</td>
<td>1.12</td>
<td>0.29</td>
<td>0.17</td>
<td>0.3</td>
</tr>
<tr>
<td>ΔTAS×P Treat</td>
<td>1,19</td>
<td>2.62</td>
<td>0.12</td>
<td>3.16</td>
<td>0.09</td>
<td>0.32</td>
<td>0.27</td>
</tr>
</tbody>
</table>

^aΔcomb was calculated as the difference between the final and initial comb area, corrected for the initial comb area.
^bT Treat = males, sham implanted (T−) versus testosterone implanted (T+) males.
^cP Treat = parasite treatment, categorical: P− males: not challenged; P+ males: challenged with infective T. tenuis larvae.
^dΔMDA was calculated as the difference in MDA concentration between S2 and S1 corrected for MDA concentration at S1.
^eΔTAS was calculated as the difference in TAS concentration between S2 and S1 corrected for TAS concentration at S1.
on metabolism 12–16 days after infection (Delahay et al. 1995) explaining this delayed effect. The parasite challenges did not reduce ornamentation in T+ males. This may be because the implants forced males to circulate high testosterone levels, which were similar in T+P− and T+P+ males, such that the exogenous testosterone would have prevented a possible parasite-induced reduction in testosterone levels and ornamentation.

Testosterone implants increased oxidative damage, as indexed by MDA concentrations (prediction 1). This might be because testosterone increased metabolic rates (e.g. Buchanan et al. 2001), or impaired the activity of antioxidant defences (e.g. Alonso-Alvarez et al. 2007), thereby increasing the oxidative stress. Smaller combed males suffered more oxidative damage (greater increase in MDA) than larger combed males when forced to circulate testosterone levels above their individual optima (T+ males only). This is consistent with a handicap mechanism (prediction 3), where the cost of testosterone (increased oxidative damage) would be greater for males with lesser ornamentation and a lower testosterone optimum (Zahavi 1975; Folstad & Karter 1992; Getty 2002).

Interestingly, elevated testosterone resulted in an increase in antioxidant defences as measured by the TAS assay in vitro, suggesting that the antioxidant defences were upregulated in vivo. However, this was not sufficient to prevent oxidative damage. Parasite challenges also increased oxidative damage (prediction 2) but reduced TAS when these were not increased by testosterone (in T−P+ as compared with T−P− males). Overall, parasite challenge caused more oxidative damage than experimental testosterone increase, possibly because of the contrasted effects of these manipulations on circulating antioxidant defences.

Testosterone can pose an oxidative challenge, which can be controlled by increasing investment in antioxidant defences (allocating resources towards self maintenance) but at the cost of investing these same resources to sexual defences (allocating resources towards self maintenance) and ornamentation. This provides a refinement of the ICHH (Folstad & Karter 1992), as the costs of increasing one increase, possibly because of the contrasted effects of testosterone and parasites on oxidative balance (changes in circulating antioxidants and oxidative damage) explained short- and medium-term effects on male ornamentation, showing for the first time that the ability to express a testosterone-dependent ornament is tightly related to an individual’s oxidative balance and susceptibility to oxidative stress. The extent to which testosterone and parasites cause oxidative damage depends on an individual’s ability to increase circulating antioxidant defences (by acquiring more or mobilizing stored antioxidants) and to resist parasites (the ability of its activated immune system to raise an appropriate immune response that finds the right target and at the same time avoid immunopathological damage; Råberg et al. 1998; von Schantz et al. 1999).

Antioxidants are depleted during immune responses (von Schantz et al. 1999), while increasing circulating antioxidants can reduce the negative impact of ROS/RNS on immune responses (Blas et al. 2006; Larbi et al. 2007). The increase in circulating antioxidants caused by testosterone implants increased oxidative damage, as testosterone-dependent ornaments partly rely on systemic antioxidants for pigmentation. However, whether carotenoid pigments act as antioxidants in birds in vivo has been both supported and questioned, and remains controversial (Costantini & Møller 2008). Clearly, more work is needed to better understand how testosterone can increase TAS, and the contribution of carotenoids to circulating antioxidant defences.

Our experiment showed that two key factors influencing ornamentation, testosterone and parasites, additively caused oxidative damage. Moreover, the impacts of testosterone and parasites on oxidative balance (changes in circulating antioxidants and oxidative damage) explained short- and medium-term effects on male ornamentation, showing for the first time that the ability to express a testosterone-dependent ornament is tightly related to an individual’s oxidative balance and susceptibility to oxidative stress. The extent to which testosterone and parasites cause oxidative damage depends on an individual’s ability to increase circulating antioxidant defences (by acquiring more or mobilizing stored antioxidants) and to resist parasites (the ability of its activated immune system to raise an appropriate immune response that finds the right target and at the same time avoid immunopathological damage; Råberg et al. 1998; von Schantz et al. 1999).

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APPENDIX A. TIMING OF THE EXPERIMENT, PROCEDURES AND DATA SAMPLING

<table>
<thead>
<tr>
<th>events</th>
<th>initial capture</th>
<th>first sampling (S1)</th>
<th>second sampling (S2)</th>
<th>third sampling (S3)</th>
</tr>
</thead>
<tbody>
<tr>
<td>dates</td>
<td>25 Sep ± 5 days</td>
<td>10 Oct ± 5 days</td>
<td>20 Oct ± 5 days</td>
<td>27 Oct ± 2 days</td>
</tr>
<tr>
<td>procedures</td>
<td>Purging of <em>T. tenuis</em> worms</td>
<td>experiment start hormone implants parasite challenges</td>
<td></td>
<td>experiment end</td>
</tr>
<tr>
<td>measurements&lt;sup&gt;a&lt;/sup&gt;</td>
<td><em>T. tenuis</em> parasites testosterone comb area MDA TAS</td>
<td><em>T. tenuis</em> parasites testosterone comb area MDA TAS</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

<sup>a</sup> We measured parasites at S1 (using faecal samples) and S3 (using direct worm counts) to check that the parasite purging conducted at S0 had been effective (S1) and to check that parasite challenges were effective (S3), respectively. We did not measure parasites at S2 because it takes 10–15 days for *T. tenuis* larvae to develop into measurable, egg-producing worms using faecal samples. Not all parameters (parasites, testosterone, MDA and TAS) could be measured for all individuals at each sampling time, due to lack of sample material (faecal samples or plasma), so sample size varies between treatment groups and sampling times.

REFERENCES


Mougeot, F., Martinez-Padilla, J., Perez-Rodriguez, L. & Bortolotti, G. R. 2007a Carotenoid-based coloration and...


