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Intraspecific sperm competition genes enforce post-mating species barriers in *Drosophila*

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Sexual selection and sexual conflict are considered important drivers of speciation, based on both theoretical models and empirical correlations between sexually selected traits and diversification. However, whether reproductive isolation between species evolves directly as a consequence of intrapopulation sexual dynamics remains empirically unresolved, in part because knowledge of the genetic mechanisms (if any) connecting these processes is limited. Here, we provide evidence of a direct mechanistic link between intraspecific sexual selection and reproductive isolation. We examined genes with known roles in intraspecific sperm competition (ISC) in *D. melanogaster* and assayed their impact on conspecific sperm precedence (CSP). We found that two such genes (*Acp36DE* and *CG9997*) contribute to both offensive sperm competition and CSP; null/knockdown lines both had lower competitive ability against *D. melanogaster* conspecifics and were no longer able to displace heterospecific *D. simulans* sperm in competitive matings. In comparison, *Sex Peptide* (*Acp70A*)—another locus essential for ISC—does not contribute to CSP. These data indicate that two loci important for sperm competitive interactions have an additional role in similar interactions that enforce post-mating reproductive isolation between species, and show that sexual selection and sexual isolation can act on the same molecular targets in a gene-specific manner.

1. Introduction

Substantial evidence suggests that sexual selection and conflict could be powerful drivers of speciation. Empirically, sister species are often differentiated by traits thought to evolve by sexual selection or conflict, and these traits are correlated with different rates of macroevolutionary diversification in a broad range of organisms [1–3]. Theoretically, models of speciation by sexual selection and sexual conflict suggest that traits involved in intrapopulation sexual dynamics can ultimately contribute to reproductive isolation [4,5]. Nonetheless, for intraspecific sexual selection to directly drive speciation requires that these two processes are mechanistically linked, but evidence of shared genetic mechanisms connecting these processes is limited [6]. Recent studies have uncovered quantitative trait loci (QTL) that control intraspecific mating differences as well as interspecies mating cues [7,8], but without knowledge of the underlying genes, the specific connection between these intraspecific mating traits and interspecific isolating barriers is unclear. As a result, whether reproductive isolation between species evolves directly as a consequence of intrapopulation sexual dynamics remains unresolved [9,10]. Here, we examine evidence for a direct mechanistic connection between intraspecific sperm competition (ISC) and post-copulatory interspecific reproductive isolation, via shared underlying genes.

Seminal fluid proteins associated with male success in sperm competition (hereafter called ‘intraspecific sperm competition’ or ‘ISC’ genes) in *Drosophila melanogaster* are among the best-characterized examples of molecules involved in sexual selection and sexual conflict. Their underlying genes are typically expressed in the male accessory gland (though other expression patterns have been observed; see [11,12]); however, they are known to influence female

oviposition rate, female remating rate and female lifespan [13], and evidence for male \times female genetic interactions at these loci is consistent with coevolution between the sexes [14]. Moreover, many of these seminal fluid proteins evolve rapidly, suggesting strong selection imposed by sexual selection and/or sexual conflict [15,16]. Rapidly evolving genes could result in elevated protein divergence between species, leading to reduced effectiveness of these proteins in heterospecific matings. These genes would then have larger relative roles in mediating reproductive isolation. Alternatively, rapid evolution could also represent strong selection to increase efficiency of sperm competition within a species, resulting in these genes having greater effects on outcompeting other sperm, regardless of whether they are conspecific or heterospecific. Given the role of sperm competition genes in intraspecific sexual interactions and their frequently rapid evolutionary rates, these genes might also be important for the development and expression of reproductive barriers between populations [17], particularly post-mating pre-zygotic reproductive barriers that are mediated by sperm competition and/or male–female interactions.

One such barrier is conspecific sperm precedence (CSP). CSP in *Drosophila* is the observation that when a female is inseminated by both conspecific and heterospecific males, the conspecific male sires the majority of the progeny, regardless of whether he is the first or second male to mate [18]. Therefore, CSP acts as a species barrier where post-mating interactions reduce the production of heterospecific (hybrid) individuals in favour of conspecific offspring. CSP is broadly observed among *Drosophila* species and other insects [19], and we confirmed its operation in crosses between *D. melanogaster* and *D. simulans* here. However, unlike genes involved in intraspecific sexual interactions, no specific genes have been identified for CSP.

Our goal in this study was to evaluate whether known ISC genes have a mechanistic role in CSP. Although there are hundreds of seminal fluid proteins, only a handful have been phenotypically characterized. We focused on three of the best-characterized genes: *Sex Peptide* (*Acp70A*) has been called a master regulator of female reproduction [20]; it is necessary for the efficient release of sperm from storage [21] and is responsible for the long-term mating response in females (i.e. reduced female receptivity, increased sperm release and oviposition [22,23]). *CG9997* is required for the transfer of three other accessory gland proteins that localize and bind *Sex Peptide* to sperm, and when lacking also results in improper release of sperm from storage [24,25]. *Acp36DE* is known to contribute to male reproductive success via effects on correct sperm storage [26,27]. We confirmed the effects of these loci on ISC and then tested these genes for a role in CSP.

2. Material and methods

(a) Fly stocks and maintenance

All flies were maintained at room temperature under standard laboratory conditions. Flies were cultured in 8 dram glass vials on Bloomington media recipe food. Wild-type female *D. melanogaster* were from the Austria w132 line, originally collected by Christian Schlötterer and donated to us by Kristi Montooth (Indiana University). Green fluorescent protein (GFP)-marked *D. melanogaster* males and *D. simulans* males were ordered from the Bloomington *Drosophila* Stock Center (32170) and

University of California Stock Center (14021-0251.263), respectively. GFP can be visualized in the pseudo-pupil and ocelli of both strains, and is a dominant marker. The creation of these lines is described by Holtzman *et al.* [28]. Null/knockdown lines for *Sex Peptide*, *CG9997* and *Acp36DE* (and control *Acp36DE*) were generously provided by Mariana Wolfner (Cornell University). For *Sex Peptide*, null males and wild-type males were generated by crossing the *SP* null line (0325/*TM3, Sb ry*) to a deficiency line (Δ 130/*TM3, Sb ry*) [22], resulting in null (0325/ Δ 130) and control (0325/*TM3, Sb ry* or Δ 130/*TM3, Sb ry*) siblings. For *Acp36DE*, we used a null *Acp36DE*¹ and control *Acp36DE*⁺. The null and control chromosomes are maintained by backcrossing every generation to *CyO/Df(2L)H2O* females (a deficiency that lacks the *Acp36DE* locus) [26]. This backcrossing generated null (*Acp36DE*¹/*Df(2L)H2O*) and control (*Acp36DE*⁺/*Df(2L)H2O*) males. For *CG9997*, we crossed a *sympUAST-CG9997* line to *tubulin-GAL4/TM3, Sb* flies to generate the knockdown males (*tubulin-GAL4/UAS-CG9997-UAS*) and wild-type males (*TM3, Sb/UAS-CG9997-UAS*) [25]. The *sympUAST-CG9997* line was donated by Mariana Wolfner, and the *tubulin-GAL4* line was ordered from the Bloomington *Drosophila* Stock Center (5138). The *CG9997* RNAi construct was previously determined to be efficient [24,25]. Because our results reiterated the described phenotypic effect of RNAi (reduced sperm competitive ability in conspecific crosses, specifically in the RNAi line but not control line), we did not further test the efficacy of RNAi for this locus.

(b) Competitive mating experimental procedure

We completed a minimum of 15 replicates of each (conspecific-GFP or heterospecific-GFP) \times (null/knockdown or control) combination for a total of 60 replicates for each locus. We focused on offensive sperm competitive ability for each locus because, after mating with a conspecific *D. melanogaster* male, *D. melanogaster* females strongly reject heterospecific *D. simulans* males; this behaviour precludes an analysis of crosses where *D. simulans* is the second male (i.e. defensive ability in *D. melanogaster*)

- (i) Day 0: virgin *D. melanogaster* females were collected and maintained in groups of 5 to age for 3–5 days prior to the first mating. This ensures that they are reproductively mature and will readily mate, and represents the beginning of the experiment. Experimental null/knockdown males, control males and GFP males were also collected as virgins and stored in the groups of 5 for 3–5 days.
- (ii) Day 1: five *D. melanogaster* females were transferred to a new vial with five GFP males (either *D. melanogaster* or *D. simulans* depending on whether the treatment was ISC or CSP, respectively) without anaesthesia and allowed to mate for 3 days. This design maximizes the number of heterospecifically mated females for use in the remaining experiment. In preliminary data collection, it was observed that mass matings resulted in a higher number of mated females compared with single-pair matings. In addition, shorter mating times resulted in very poor average mating success, owing to premating behavioural isolation between the species.
- (iii) Day 4: females were transferred individually to a fresh vial of food and allowed to oviposit for 1 day. In general, *D. melanogaster* females refuse to remate within 24 h [29,30], so isolating females increases the chances that they will readily mate with the second male. Isolating females for longer periods of time can result in use of all the sperm from the first male, and the effect we would observe would not be due to sperm competition. Females that did not lay eggs when isolated for one day were discarded, as these females are assumed not to have mated with the GFP-marked male.
- (iv) Day 5: females were transferred to a fresh vial, to which a single experimental or control male was added. (In

preliminary data collection, we observed matings at this time point and found that all females had mated with the second male within 2 h, regardless of genotype; therefore, we did not continue to watch matings for the remainder of the experiment). These pairings were maintained and transferred daily until day 7, when all flies were discarded. Daily transfers reduce crowding in a given vial, so that hybrid progeny will not be absent owing to larval competition. After adults were removed, cotton substrate was added to day 5–7 vials to encourage pupation of hybrid individuals because *D. simulans* (and to some extent hybrids) make pupal mats rather than climb up the side of the glass vials.

- (v) Scoring progeny: vials of days 4–7 were kept until all progeny had eclosed. As progeny began to emerge they were sexed and scored under UV light for the presence of the GFP (visible in pseudo-pupil and ocelli). The phenotype used in the data analysis is the proportion of GFP progeny in vials from day 5 to day 7. For conspecific matings we used the total number of progeny and for heterospecific matings, we restricted the proportion to the number of females, because male hybrids are inviable (results were not different if we used proportion of females for both CSP and ISC assays). We excluded any replicates where we could not ensure that the first male had mated (i.e. where no progeny were observed in the day 4 vial when the female was isolated after first mating) or that the second male had mated (i.e. where all progeny were GFP). Within these constraints, we were still able to maintain a sample size of 15 replicates per treatment. To confirm that CSP operates between *D. melanogaster* and *D. simulans*, we also performed this experiment (with the same replication and quality controls) using wild-type *D. simulans* as the first male and *D. melanogaster*-GFP as the second male.

(c) Statistical analysis

Differences in competitive ability between null and wild-type males were compared using a logit model in R:

$$\text{logit}(p_i) = \beta_0 + \beta_1 \times x_1 + \beta_2 \times x_2 + \beta_3 \times x_1x_2.$$

The binomial response variable was the number of green-eyed progeny observed compared with the number of wild-type progeny observed, as this represents the magnitude of the sperm competitive ability of the first male versus the second male, regardless of whether the first male is conspecific or heterospecific. The variable x_1 is a binary variable describing whether the second male to mate was null/knockdown or wild-type ($x_1 = 0$ for wild-type males), and β_1 is the log-odds describing the difference in competitive ability between null/knockdown and wild-type males. The variable x_2 is a binary variable describing whether the GFP-marked male is conspecific or heterospecific ($x_2 = 0$ for *D. melanogaster*-GFP males) and β_2 is the log-odds for the difference in competing against conspecific versus heterospecific males. The last variable is also a binary variable, describing the interaction between the first two variables ($x_1x_2 = 1$ for null/knockdown males competing with *D. simulans* GFP males), and β_3 describes the log-odds for the interaction term.

The differences in the total number of progeny produced between crosses that involved control males and crosses that involved null/knockdown males were compared using a linear model similar to the model above in R. In this model, the response variable was the total number of progeny produced after the second mating, and we were primarily interested in the significance of β_1 (which we refer to as β_{null} to reduce redundancy). The differences in oviposition and total number of progeny produced after mating with only a control or null genotype for *Acp36DE* over 3 days (analogous to days 5–7

described above, but without a first mating having had occurred) were compared using a Wilcoxon rank-sum test in R. For these data, we used eight replicates for each genotype and counted eggs in each vial after the female and male pair were transferred without aspiration to a new vial.

(d) Measures of molecular evolution

Nucleotide alignments of the coding regions (CDS) for all genes identified as members of the *Sex Peptide* network [31] and *Acp36DE* were downloaded from UCSC genome browser. The species and species tree we used in the analysis were (*D. melanogaster* (*D. simulans*, *D. sechellia*)). A single d_N/d_S value was estimated for the entire unrooted tree using PAML (M0 model) [32]. Besides providing the phylogeny, all other options were left as default. Pairwise comparisons of the total number of non-synonymous and synonymous changes were made for all genes using the Nei–Gojobori method for just *D. melanogaster* and *D. simulans*. This was completed using MEGA v. 6 [33].

3. Results and discussion

Using sperm-competition assays, we assessed the sperm competitive ability of null/knockdown males versus wild-type individuals at each locus (figure 1). Sperm competition assays involve mating a female sequentially to males of two different genotypes, to determine the ability of a given male genotype either to resist displacement by the second male (defensive ability) or to displace the first male's sperm (offensive ability); competitive ability is revealed in the relative siring success of each male. This same design can be used to assess both ISC and CSP, simply by varying whether the first mating involves conspecific or heterospecific males, respectively (figure 1). In our experiments, either null/knockdown or wild-type (control) males were the second males to mate following a first mating with either a wild-type (GFP) conspecific male (to confirm a role in ISC) or a wild-type heterospecific (GFP) male (to assess CSP). Our assays therefore assess the offensive ability of each null/knockdown male in comparison with conspecific wild-type males. Using GFP-marked first males allows us to quantify progeny siring rates directly in offspring using this dominant marker.

We found that two ISC loci also had large and significant effects on the magnitude of CSP (figure 2). *Acp36DE* is known to contribute to both defensive and offensive ability; in our offensive assay, null males were less successful competitors than wild-type males when competing against conspecific males, confirming previous results ($\beta_1 = 0.9409$, $p < 0.0001$; figure 2a). Importantly, null males were also less successful than wild-type males when competing against heterospecific males ($\beta_2 = -2.9463$, $p < 0.0001$; figure 2a). The effect of *Acp36DE* on CSP was also greater than the effect on ISC ($\beta_3 = 0.7644$, $p < 0.0001$); that is, the magnitude of the difference between null versus wild-type males is greater when measuring interspecific than ISC. Similar to *Acp36DE*, *CG9997* contributed both to ISC and CSP. When competing against conspecific males, knockdown males were less successful at displacing sperm than wild-type males ($\beta_1 = 0.3465$, $p < 0.0001$; figure 2b). This pattern was also seen for knockdown versus wild-type males when competing against heterospecific males ($\beta_2 = -1.7399$, $p < 0.0001$), and again the magnitude of the effect is greater for CSP compared with ISC ($\beta_3 = 0.4205$, $p = 0.0064$; figure 2b).

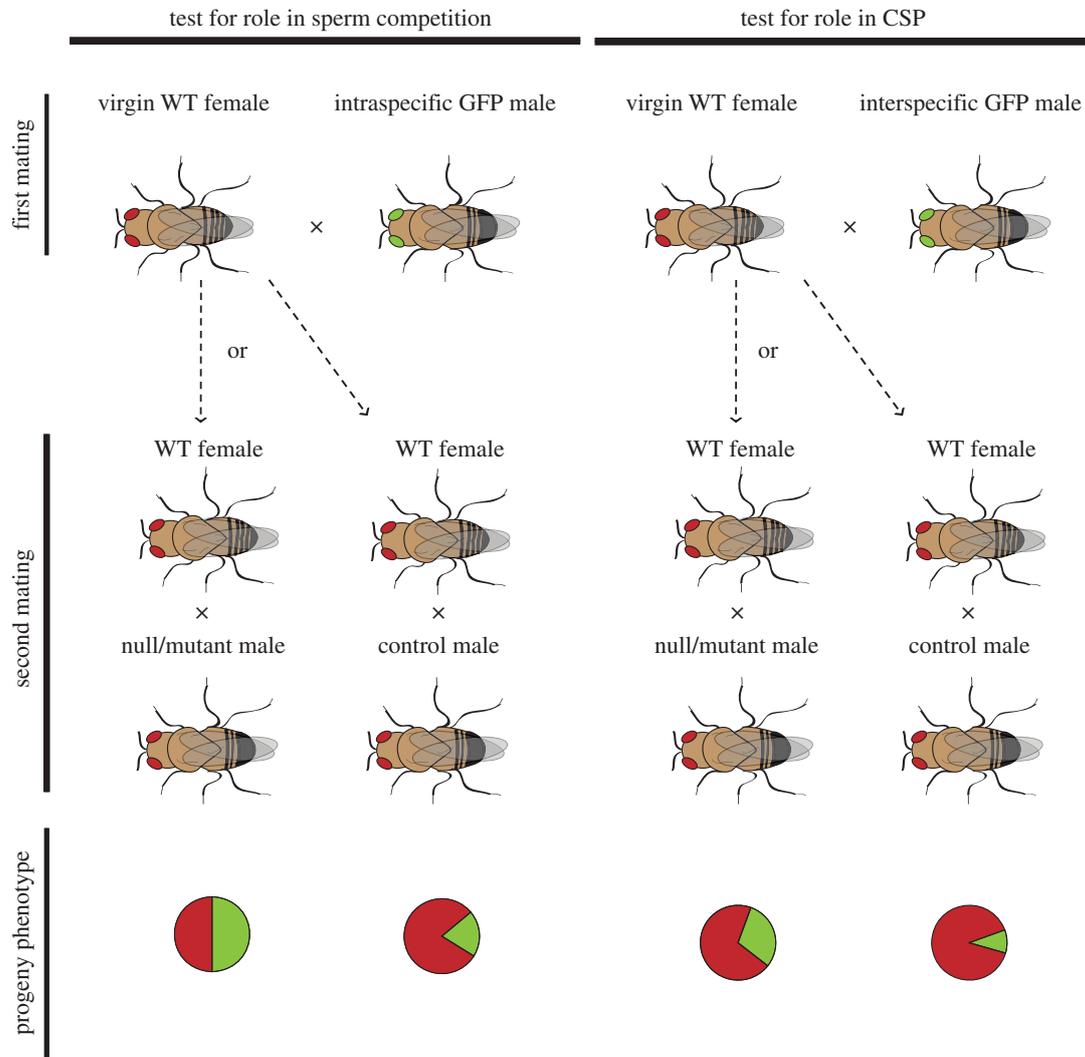


Figure 1. Experimental design of the sperm competition assay. The same design is used to test for a role in ISC and CSP, by varying the identity of the first mated (GFP) male. The response phenotype is the proportion of progeny with green eyes; schematic expectations for each case are represented with pie charts. For genes involved in ISC (left-hand side), females are expected to produce proportionately more GFP offspring when mated to null/knockdown second males, compared with wild-type second males, as these individuals are less able to displace first male (GFP) sperm (owing to reduced offensive competitive ability). For genes involved in CSP (right-hand side), the same pattern is expected to hold, although the total proportions of green-eyed progeny are expected to be less than the respective crosses assessing ISC because of reduced compatibility in heterospecific crosses. (Online version in colour.)

Unlike these two loci, *Sex Peptide* did not affect the magnitude of CSP, although it had an effect on sperm offensive ability among conspecifics (figure 2c). Compared with wild-type males, null *Sex Peptide* males were less successful at displacing conspecific sperm ($\beta_1 = 0.9556$, $p < 0.0001$). By contrast, wild-type and null males both successfully displaced sperm from heterospecific males ($\beta_2 = -1.5036$, $p < 0.0001$ and $\beta_3 = -0.9612$, $p < 0.0001$, respectively; figure 2c). Although *Sex Peptide* is better known for its defensive role in conspecific matings [14,21], which we also confirmed here (electronic supplementary material, figure S1), our analysis shows that it also plays a clear role in offensive ISC, but does not contribute to CSP-mediated reproductive isolation through offensive ability.

Differences in fertility or in differences in mating frequency/copulation intensity between control and null/knockdown males could result in spurious observation of differences in competitive ability. We evaluated whether control and null/knockdown males differed in fecundity by comparing the total number of progeny produced for the competitive crosses containing control males versus null/

knockdown males (electronic supplementary material, figures S2–S4). For *CG9997* and *Sex Peptide*, there was no difference in the number of progeny produced between control and null crosses ($\beta_{\text{null}} = 11.467$; $p = 0.307$ for *CG9997*, and $\beta_{\text{null}} = -1.867$; $p = 0.866$ for *Sex Peptide*). For *Acp36DE*, crosses involving the null male did produce fewer progeny ($\beta_{\text{null}} = -26.628$; $p = 0.0129$), similar to previous reports that also analysed sperm competitive ability for this locus [26], but this reduction was observed regardless of whether this male was competing with a heterospecific or conspecific male. Because we could not disentangle fecundity effects from competitive effects for *Acp36DE*, we also looked at differences in oviposition and progeny production over 3 days after single (non-competitive) matings with either control or null genotypes for this locus. This time frame is identical to the time frame of our experimental design. We observed no difference between number of eggs laid over the 3 days ($W = 31.5$; $p = 1.00$) or progeny produced ($W = 35$; $p = 0.751$) between the null and control *Acp36DE* male genotypes (electronic supplementary material, figures S5–S6).

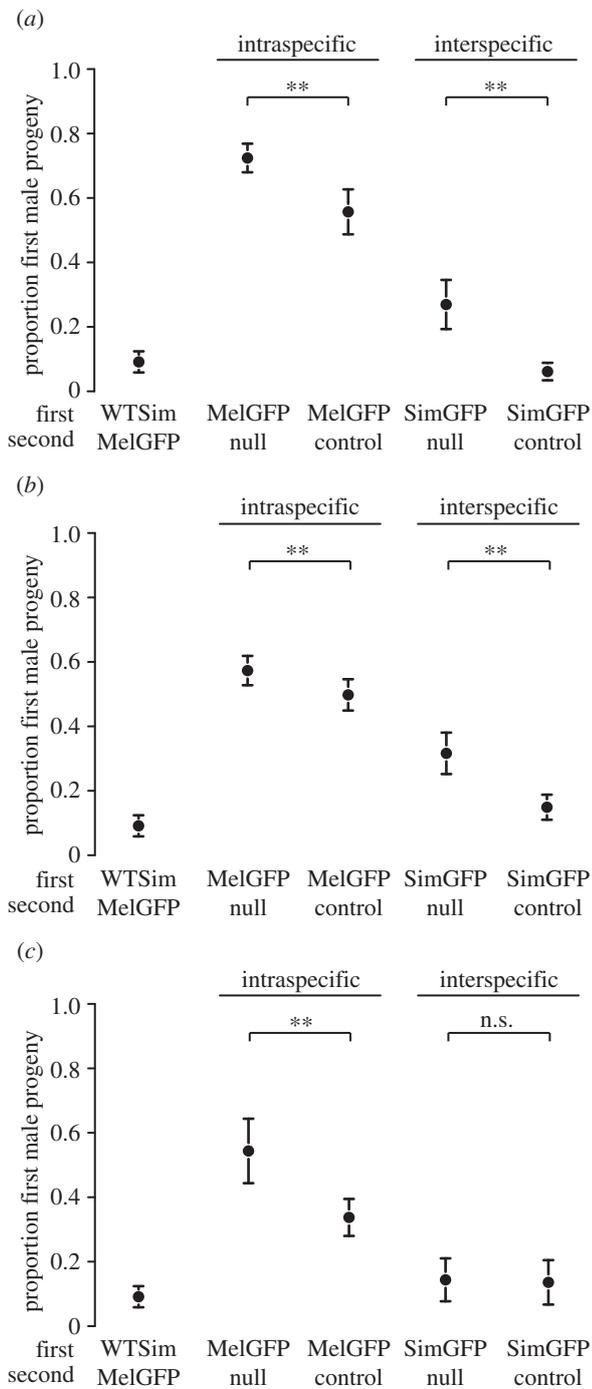


Figure 2. The ability of null/knockdown versus wild-type males to displace competitor sperm when the competitor is either conspecific or heterospecific for each locus we examined. (a) *Acp36DE*, (b) *CG9997* and (c) *Sex Peptide*. Every panel depicts the control: wild-type *D. simulans* competing against GFP *D. melanogaster*. Each point represents mean \pm s.e. proportion of 15 replicates. Asterisk designates a null/knockdown versus wild-type male comparison that is significant from the logistic regression. n.s. stands for not significant.

If mating frequency and intensity is correlated with progeny production, these data also suggest that there was no difference in mating frequency between null/knockdown and control males. Interestingly, previous studies have demonstrated that females that mate with *Sex Peptide* null males remate more quickly in comparison with females initially mated to control males [23]; remating rate might therefore be increased using null *Sex Peptide* males. Because *CG9997* is similarly thought to play a role in remating rate and the long-term response, similar logic could also apply

for this locus. In either case, this would result in increased matings by null/knockdown males compared with control males, which could not explain why null/knockdown males specifically performed poorly in CSP assays. We therefore infer that differences in fertility and mating frequency of control versus null/knockdown males are unlikely to explain the observed involvement of *CG9997* and *Acp36DE* in CSP.

To examine the evolutionary dynamics of *Acp36DE*, *CG9997* and *Sex Peptide*, we examined sequence data for each locus and for 11 additional genes known to be involved in the *D. melanogaster* 'sex peptide network' of which *Sex Peptide* and *CG9997* (but not *Acp36DE*) are known members [31]. Of the 14 genes, our three focal loci had relatively high values of d_N (the number of non-synonymous substitutions per non-synonymous site; table 1), reflective of high relative rates of protein-changing evolution. However, there was no obvious distinction between the values of d_N observed at the two genes found to affect CSP (*CG9997* and *Acp36DE*) and the one that did not (*Sex Peptide*; table 1). Likewise, although the values of d_N/d_S (which can provide evidence for positive selection between species when d_N/d_S is >1) were uniformly higher in all 14 loci compared with the *D. melanogaster* (*D. simulans*) genome-wide average of 0.07 (0.11) [34], these estimates of protein-coding evolution do not clearly differentiate *CG9997* and *Acp36DE* from *Sex Peptide* (table 1).

Overall, our analyses support several new findings. First, they demonstrate that *CG9997* and *Sex Peptide* play a role in offensive sperm competition (previous analyses of these loci only focused on defensive sperm competition). Second, they uncover two genes that are mechanistically involved in CSP. Although studies have previously mapped QTL for CSP [35,36], no genes have yet been identified for this widespread post-mating pre-zygotic reproductive isolating barrier. (Note that neither *Acp36DE* nor *CG9997*, nor any other 'male gene' members of the Sex Peptide network, colocalize with the described CSP QTL in *Drosophila* [36]). Second, because *Acp36DE* and *CG9997* are known actors in sperm competition and sperm storage within *D. melanogaster*, their involvement in CSP provides evidence for a direct mechanistic connection between conspecific and heterospecific competitive sexual interactions, via the same causative loci. Previous studies have linked sexually selected traits with reproductive isolation by demonstrating that specific phenotypes are associated both with sexual selection and reproductive isolation [37,38], although the genetics of these traits remain unknown. Other studies have deduced that genomic regions (QTL) associated with reproductive isolation are also sexually selected; however, the contribution of sexual selection is inferred (generally via assortative mating), rather than demonstrated directly [39,40]. Nonetheless, for models of speciation via sexual selection to be plausible, at least one locus must be demonstrably involved in both sexual selection and reproductive isolation. Here, we have identified two such loci.

Although our data show that specific molecular players can mediate both ISC and CSP, not all ISC genes appear to be involved in CSP, at least via the mechanisms we probably observed here (sperm storage, retention and release from storage). (Note, however, that these genes could still contribute to CSP through their effects on female remating rate, particularly as remating sets the stage for CSP.) Differences among genes observed here could be explained by differences in their evolutionary rates of change, and/or in the magnitude

Table 1. Estimates of d_N , d_S , and d_N/d_S for genes in the sex peptide network and *Acp36DE* using *D. melanogaster*, *D. simulans* and *D. sechellia*, with the number of codons, non-synonymous changes (N) and synonymous changes (S) from a pairwise comparison of *D. melanogaster* and *D. simulans*. Asterisk denotes genes examined in this study.

gene	chromosome position	d_N	d_S	d_N/d_S	codons	N	S
<i>CG1652</i>	chr2R:5703015–5704105	0.1258	0.4172	0.3014	323	90.083	58.917
<i>CG3239</i>	chrX:5387007–5390082	0.0887	0.2618	0.3387	842	101	71
<i>Acp36DE*</i>	chr2L:18356047–18359026	0.0831	0.1697	0.4895	913	95.5	79.5
<i>Sex Peptide*</i>	chr3L:13294735–13295022	0.0599	0.2228	0.2686	56	4	4
<i>CG30488</i>	chr2R:8734353–8735509	0.0491	0.2205	0.2226	266	28.167	27.833
<i>CG12558</i>	chr3R:24616244–24617183	0.0422	0.1804	0.2338	299	17	25
<i>CG9997*</i>	chr3R:24581206–24582701	0.0382	0.1196	0.3191	331	26	20
<i>Seminase</i>	chr3L:20948855–20949850	0.0374	0.1135	0.3296	276	21	18
<i>CG5630</i>	chr3R:16740018–16754935	0.0330	0.1910	0.1727	378	26.5	31.5
<i>CG14061</i>	chr3R:24582721–24583956	0.0258	0.1841	0.1403	367	17	33
<i>CG1656</i>	chr2R:5704805–5706040	0.0177	0.1390	0.1272	329	10.5	17.5
<i>Esp</i>	chr3R:20667657–20676134	0.0048	0.1530	0.0315	655	5	50
<i>CG17575</i>	chr2R:8733049–8734101	0.0040	0.1991	0.0219	292	1	21
<i>SPR</i>	chrX:5340689–5388045	0.0039	0.3281	0.0117	436	4	59

of pleiotropy experienced by these genes. In the first case, genes that are more rapidly evolving could be more likely to contribute to CSP because they have increased functional divergence between species. However, we find no clear differences among our three focal genes in common measures of protein evolution (table 1), suggesting that differences in the rates of protein evolution *per se* are not responsible for differential involvement of these ISC genes in CSP. In the second case, genes with smaller pleiotropic effects might be less constrained, therefore more responsive to selection for increased reproductive isolation, and correspondingly more important for CSP. Although all three genes studied here are required for effective sperm competition, *Sex Peptide* is known to be involved in at least nine different phenotypes [20], whereas *Acp36DE* is only known to act on sperm accumulation [26], and *CG9997* is only known to be required for the transfer of certain accessory gland proteins into females [25]. Therefore, current data suggest that the locus without an effect on CSP (*Sex Peptide*) is a highly pleiotropic gene, whereas the other two loci are less pleiotropic by comparison, possibly contributing to the differential involvement of these genes in CSP.

Finally, if the functional significance of some but not all post-mating pre-fertilization mechanisms differs in the context of sexual selection versus reproductive isolation, this could explain the differential involvement of specific sperm competition genes in CSP. Our observations that *Acp36DE* and *CG9997* clearly contribute to both ISC and CSP indicate that their functional roles in proper sperm storage and utilization [24,26] are important mechanisms for both ISC and CSP. This inference is consistent with recent studies in *D. melanogaster*, *D. simulans* and *D. mauritiana* that suggest that siring success in both ISC and CSP is determined by female control of sperm use and displacement, although the specific patterns of use and displacement are different in the two reproductive contexts [41,42]. By contrast, our finding that *Sex Peptide* does not contribute to differences in siring success in CSP indicates that the specific functional roles of this

critical ISC gene are less important in the altered reproductive context of CSP. Our findings might also suggest additional, previously undescribed functional roles for *CG9997* and *Acp36DE*, specifically in the context of CSP. For example, the known role of *CG9997* is to transfer other seminal fluid proteins into the reproductive tract, which then bind *Sex Peptide* to sperm [25]. *Sex Peptide* and *CG9997* are therefore mechanistically linked, and they both show similar sperm release phenotypes in intraspecific matings [25,21], which is consistent with our ISC results. However, our finding that *CG9997*, but not *Sex Peptide*, has a phenotypic effect on CSP suggests that *CG9997* or the proteins it helps transfer have additional roles not directly linked to the function of *Sex Peptide*, at least in the alternative reproductive context of interspecific matings. These inferences can be more directly tested with further functional data in the future.

4. Conclusion

Seminal fluid loci are strong *a priori* candidates for genes that could play dual roles in intraspecific sexual interactions and interspecific reproductive isolation. They are critical for sexual selection, frequently evolve rapidly and have functions that appear to be phenotypically and behaviourally similar to those operating during post-mating pre-zygotic sexual isolation. Although it remains to be seen how many other sperm competition proteins are similarly involved in species reproductive barriers, our data confirm that at least two known *D. melanogaster* seminal fluid proteins do have a second function in sexual isolation. Accordingly, these findings confirm that a necessary condition for sexual selection/conflict to drive sexual isolation—a shared genetic basis for these processes—is met for ISC and CSP. Further studies linking these two processes will allow us to better understand how reproductive isolation is facilitated or constrained by sexual selection, and which loci are unique versus shared between these processes. Although sexual selection and sexual conflict

are often discussed as important drivers of speciation, this broader understanding of the prevalence of direct genetic connections between these processes is essential for evaluating their relative importance compared with other drivers of isolation and diversification.

Data accessibility. All data used in the study are available as an electronic supplementary material file.

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